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## **Data Sheet**

### **PDE7A/CRE Reporter - HEK293 Cell Line Catalog #: 60413**

#### **Description**

Phosphodiesterases (PDEs) regulate the intracellular levels of cAMP and cGMP by hydrolyzing cAMP and cGMP to their inactive 5' monophosphates. These cyclic nucleotides play an important role as second messengers in diverse physiological functions. PDE7 is a cAMP-specific enzyme and two PDE7 genes (PDE7A and PDE7B) have been identified. PDE7A is widely expressed in various tissues including skeletal muscle, T lymphocytes, brain and pancreas. Inhibition of PDE7 activity by its inhibitors leads to elevated intracellular level of cAMP.

The regulation of intracellular level of cAMP by PDE7A in cells can be monitored by a specific reporter for the cAMP pathway, the CRE luciferase reporter. CRE luciferase reporter contains a luciferase gene that is under the control of the cAMP response element (CRE). Elevation of intracellular cAMP activates cAMP response element binding protein (CREB) to bind CRE and induces the expression of luciferase.

The PDE7A/CRE Reporter-HEK293 line contains the firefly luciferase gene under the control of CRE as well as a constitutive expression construct for human PDE7A (phosphodiesterase 7A, accession number NM\_002603), both stably integrated into HEK293 cells. The luciferase expression level from the CRE reporter is used to monitor the activity of PDE7A in the cells. The cell line is validated for the induction of the expression of luciferase reporter by an inhibitor of PDE7A.

#### **Application**

- Monitor human PDE7A activity.
- Screen for PDE7A inhibitors.

#### **Format**

Each vial contains  $\sim 2 \times 10^6$  cells in 1 ml of 10% DMSO.

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### **Functional Validation and Assay Performance**

N-terminal FLAG-tagged human PDE7A is stably expressed in a human embryonic kidney (HEK293) cell line. PDE7A expression was confirmed by Western blotting.

PDE7A activity was monitored by CRE luciferase activity assay. Forskolin is commonly used to raise the intracellular level of cAMP in cell physiology studies. Forskolin resensitizes cell receptors by activating the enzyme adenylyl cyclase and increasing cAMP levels. When cells were activated by forskolin, the level of cAMP was upregulated in control HEK293 cells containing only CRE luciferase reporter (CRE Reporter-HEK293) inducing high expression of the CRE luciferase reporter. However, PDE7A/CRE Reporter-HEK293 cells showed reduction in the level of forskolin-induced cAMP, resulting in lowered expression of luciferase. Inhibition of PDE7A activity by BRL 50481, a PDE7A inhibitor, restored the cAMP level, resulting in higher luciferase activity (See figure 1., below).

### **Culture Conditions**

**Thaw Medium 1 (BPS Cat. #60187):** MEM/EBSS (with L-glutamine) (Hyclone #SH30024.01) medium supplemented with 10% FBS (Life technologies #26140-079), 1% non-essential amino acids (Hyclone #SH30238.01), 1 mM Na pyruvate (Hyclone #SH30239.01), 1% Penicillin/Streptomycin (Hyclone SV30010.01)

**Complete Growth Medium:** Thaw Medium 1 (BPS Cat. #60187), plus 400 µg/ml of Geneticin (Life technologies #11811031) and 50 µg/ml of Hygromycin B (Hyclone #SV30070.01).

Cells should be grown at 37°C with 5% CO<sub>2</sub> using complete growth medium (Thaw Medium 1, Geneticin, and Hygromycin B).

It may be necessary to adjust the percentage of CO<sub>2</sub> in the incubator depending on the NaHCO<sub>3</sub> level in the basal medium.

**To thaw the cells,** it is recommended to quickly thaw the frozen cells from liquid nitrogen in a 37°C water-bath, transfer to a tube containing 10 ml of Thaw Medium 1 (**no Geneticin and Hygromycin**), spin down cells, and resuspend cells in pre-warmed Thaw Medium 1 (**no Geneticin and Hygromycin**). Transfer resuspended cells to a T25 flask and culture in a CO<sub>2</sub> incubator at 37°C overnight. The next day, replace the medium with fresh Thaw Medium 1 (**no Geneticin and Hygromycin**), and continue growing culture in a CO<sub>2</sub> incubator at 37°C until the cells are ready to be split. At first passage, switch to complete growth medium (**contains Geneticin and Hygromycin**). Cells should be split before they reach complete confluence.

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**To passage the cells**, rinse cells with phosphate buffered saline (PBS), and detach cells from the culture vessel with 0.05% Trypsin/EDTA. Add complete growth medium and transfer to a tube, spin down the cells, then resuspend cells and seed appropriate aliquots of cell suspension into new culture vessels. Subcultivation ration: 1:10 to 1:20 weekly or twice a week.

**Assay performance: Induction of CRE reporter activity by PDE7A inhibitor in PDE7A /CRE Reporter – HEK293 cells**

The following assays are designed for 96-well format. To perform the assay in different tissue culture formats, the cell number and reagent volumes should be scaled appropriately.

**Materials Required but Not Supplied**

- BRL50481 (Enzo # BML-PD120): PDE7A inhibitor. Prepare 100 mM of stock solution in DMSO.
  - Forskolin: prepare 10 mM of stock solution in DMSO.
  - Assay medium: Thaw Medium 1 (BPS Cat. #60187)
  - 96-well tissue culture plate or 96-well tissue culture-treated white clear-bottom assay plate (Corning # 3610)
  - Luciferase reagents for measuring firefly luciferase activity (We use ONE-Glo™ luciferase assay system, Promega # E6110. Other luciferase assay systems are also suitable).
  - Luminometer
1. Harvest cells from culture in Growth medium and seed cells at a density of ~30,000 cells per well in 40µl of Thaw Medium 1 in a white clear-bottom 96-well microplate.
  2. Dilute PDE7A inhibitor (BRL50481) stock in Thaw Medium 1. Add 10 µl of diluted inhibitor to wells. The final DMSO concentration in our assay is 0.1% (the final DMSO concentration may be up to 0.5%).

Add 10 µl of Thaw Medium 1 containing the same concentration of DMSO to control wells without inhibitor.

Add 50 µl of Thaw Medium 1 with 0.1% DMSO to cell-free control wells (for determining background luminescence).

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Tap the plate gently to mix. Make sure that the cells distribute evenly in the wells.

3. Incubate at 37°C in a CO<sub>2</sub> incubator for ~ 16 hours.
4. The next day, cells should be ~ 80% confluent. Dilute Forskolin in Thaw Medium 1 to 10 µM and add 5.5 µl of diluted Forskolin to stimulated wells. The final Forskolin concentration in the wells is 1 µM.

Add 5.5 µl of Thaw Medium 1 with the same concentration of DMSO to the unstimulated control wells and cell-free control wells.

Tap the plate *very gently* to mix. Make sure that cells remain attached to the wells.

Set up each treatment in at least triplicate.

5. Incubate the plate at 37°C in a CO<sub>2</sub> incubator for 5-6 hours.
6. Perform luciferase assay using ONE-Glo™ luciferase assay system: Add 50 µl of ONE-Glo Luciferase reagent per well and rock at room temperature for ~15 minutes. Measure luminescence using a luminometer. If using other luciferase reagents from other vendors, follow the manufacturer's assay protocol.
7. Data Analysis: Subtract average background luminescence (cell-free control wells) from luminescence reading of all wells.  
The fold induction of CRE luciferase reporter expression = average background-subtracted luminescence of Forskolin-stimulated wells / average background-subtracted luminescence of unstimulated control wells

**Figure 1. PDE7A overexpression reduces the level of cAMP following forskolin stimulation in PDE7A /CRE-HEK293 cells. This effect is reversed by BRL 50481, a PDE7A inhibitor.**

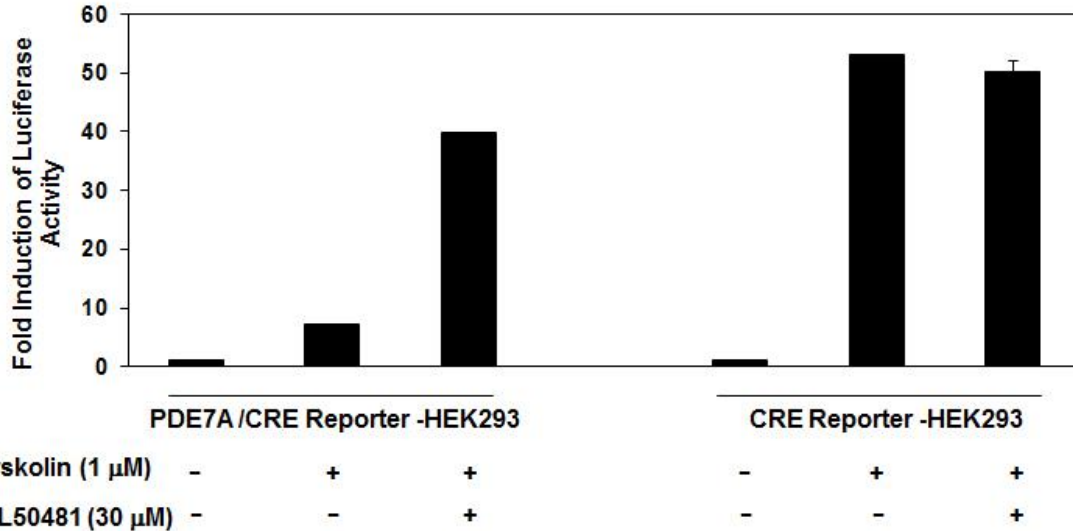
The data are shown as fold induction of CRE luciferase reporter activity. Fold induction was determined by comparing values against the mean value for control cells without forskolin treatment (fold induction = background-subtracted luminescence of Forskolin-stimulated wells / average background-subtracted luminescence of unstimulated control wells).

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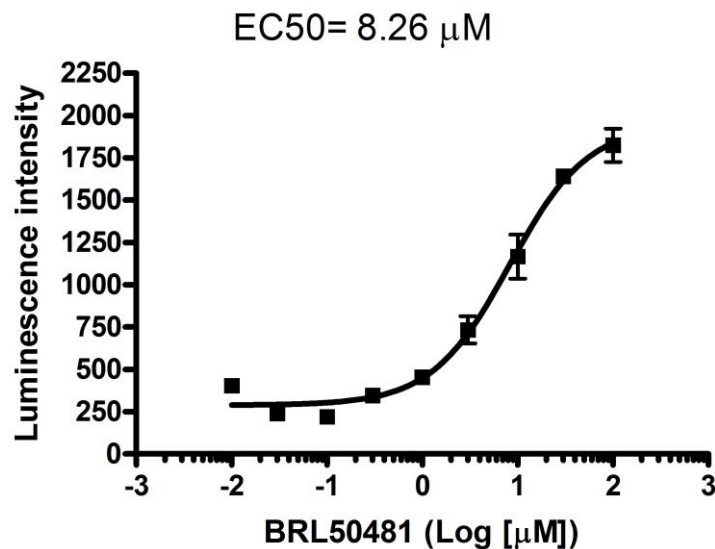
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**Figure 2. BRL50481 inhibition dose response curve in PDE7A /CRE Reporter cells.**

The result is shown as background-subtracted luminescence intensity. The inhibition of PDE7A in cells induces luminescence, so the inhibitory effects of BRL50481 on PDE7A activity is expressed as EC50. The EC50 of BRL50481 is ~ 8.26 μM.



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### **Mycoplasma testing**

The cell line has been screened using the PCR-based Venor®GeM Mycoplasma Detection kit (Sigma-Aldrich) to confirm the absence of Mycoplasma species.

### **Storage**

Store cells in liquid nitrogen upon arrival.

### **Vector and sequence**

N-terminal FLAG-tagged human PDE7A (accession number NM\_002603) was cloned into pcDNA3.1 vector (Invitrogen).

Polylinker: CMV-HindIII-KpnI-BamHI-**PDE7A**-XhoI-XbaI-ApaI-----SV40-neomycin<sup>R</sup>

hPDE7A sequence:

MDYKDDDDKGITLIWCLALVLIKWITSKRRGAISYDSSDQTALYIRMLGDVVRVRS  
RAGFESERRGSHPYIDFRIFHSQSEIEVSVSARNIRRLLSFQRYLRSSRFFRGTA  
VNSLNILDDDYNGQAKCMLEKVGWNWFDIFLFDRLTNGNSLVSLTFHLFSLHG  
LIEYFHLDMMKLRRFLVMIQEDYHSQNPYHNAVHAADVTDQAMHCYLKEPKLAN  
SVTPWDILLSLIAAATHDLDPGVNQPFLIKTNHYLATLYKNTSVLENHHWRSVA  
GLLRESGLFSLPLESRQQMETQIGALILATDISRQNEYLSLFRSHLDRGDLGLE  
DTRHRHLVLQMAKCADICNPCRTWELSKQWSEKVTSEFFHQGDIEKKYHLGV  
SPLCDRHTESIANIQIGFMTYLVEPLFTEWARFSNTRLSQTMLGHVGLNKASWK  
GLQREQSSSEDTDAAFELNSQLLPQENRLS

### **References**

1. Malik, R. *et al.* (2008) Cloning, stable expression of human phosphodiesterase 7A and development of an assay for screening of PDE7 selective inhibitors. *Appl. Microbiol. Biotechnol.* **77 (5)**: 1167-1173
2. Fan Chung, K. (2006) Phosphodiesterase inhibitors in airways disease. *Eur. J. Pharmacol.* **533(1-3)**:110-117

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### **Related Products**

<b><u>Product Name</u></b>	<b><u>Catalog #</u></b>	<b><u>Size</u></b>
CRE Reporter-HEK293 cell line	60613	2 vials
PDE7A-HEK293 Cell line	60407	2 vials
PDE7B-HEK293 Cell line	60412	2 vials
Rat PDE7A-HEK293 Cell line	60408	2 vials
CRE/CREB Kit (cAMP/PKA)	60611	500 rxns
PDE4D Cell-Based Activity Assay Kit	60505	500 rxns
PDE7A Enzyme (Human)	60070	10 µg
PDE7B Enzyme (Human)	60071	10 µg
PDE7A Enzyme (Rat)	60074	10 µg
PDE7B Enzyme (Rat)	60075	10 µg

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