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Data Sheet

Hedgehog Signaling Pathway Gli Reporter – NIH3T3 Cell Line Catalog #: 60409

Product Description

The *Gli Reporter – NIH3T3 Cell Line* is designed for monitoring the activity of the hedgehog signaling pathway. The hedgehog pathway controls stem cell growth in embryonic and adult tissues and promotes tumor growth in a number of cancers. The mammalian hedgehog proteins, Sonic Hedgehog (Shh), Indian Hedgehog (Ihh), and Desert Hedgehog (Dhh) activate hedgehog signaling by binding to their membrane receptor "Patched" (PTCH). This binding releases PTCH inhibition of Smoothened (Smo) and allows Smo to activate the Gli family of transcription factors, leading to transcription and expression of hedgehog signal target genes.

The *Gli Reporter – NIH3T3 Cell Line* contains the firefly luciferase gene under the control of Gli responsive elements stably integrated into NIH3T3 cells. Luciferase expression correlates with activation of the hedgehog signaling pathway. This cell line is validated for its response to stimulation with murine Sonic Hedgehog and to treatment with inhibitors of the hedgehog signaling pathway.

Application

- Monitor hedgehog signaling pathway activity.
- Screen for activators or inhibitors of the hedgehog signaling pathway.

Format

Each vial contains $\sim 1.5 \times 10^6$ cells in 1 ml of 10% DMSO.

Storage

Immediately upon receipt, store in liquid nitrogen.

General Culture Conditions

Thaw Medium 5 (BPS Cat. #60182): DMEM medium (Hyclone #SH30243.01) supplemented with 10% Calf Serum (CS) (Hyclone # SH30072.03), (*not* FBS), 1% Penicillin/Streptomycin (Hyclone # SV30010).

Growth Medium 5B (BPS Cat. #79541): Thaw Medium 5 (BPS Cat. #60182) and 500 μ g/ml of Geneticin (LifeTechnologies #11811031).

Cells should be grown at 37° with 5% CO₂ using Growth Medium 5B.

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To thaw the cells, it is recommended to quickly thaw the frozen cells from liquid nitrogen in a 37°C water-bath, transfer to a tube containing 10 ml of Thaw Medium 5 (**no Geneticin**), spin down cells, resuspend cells in pre-warmed Thaw Medium 5 (**no Geneticin**), transfer resuspended cells to a T25 flask and culture in a CO₂ incubator at 37°C. At first passage, switch to Growth Medium 5B (**contains Geneticin**). Cells should be split before they reach complete confluence.

To passage the cells, rinse cells with phosphate buffered saline (PBS), and detach cells from the culture vessel with 0.05% Trypsin/EDTA. Add Growth Medium 5B and transfer to a tube, spin down the cells, then resuspend cells and seed appropriate aliquots of cell suspension into new culture vessels. Subcultivation ration: 1:10 to 1:20 weekly or twice a week.

To freeze down the cells, rinse cells with phosphate buffered saline (PBS), and detach cells from culture vessel with Trypsin/EDTA. Add Growth Medium 5B and transfer to a tube, spin down cells, and resuspend in freezing medium (10% DMSO + 90% FBS). Place at -80°C overnight and place in liquid nitrogen the next day. Alternatively, vials may be placed directly in liquid nitrogen.

Functional Validation and Assay Performance

The following assays are designed for 96-well format. To perform the assay in different tissue culture formats, the cell number and reagent volume should be scaled appropriately.

Materials Required but Not Supplied

- Mouse Sonic Hedgehog (Shh), His-tag (BPS Bioscience # 91012)
- Cyclopamine (BPS Bioscience # 27013): inhibitor of hedgehog pathway (Smo inhibitor)
- Vismodegib (GDC-0449) (BPS Bioscience #27010): inhibitor of hedgehog pathway (Smo inhibitor)
- Assay medium: Opti-MEM Reduced Serum Medium (Invitrogen #31985-062) with 0.5% calf serum (Hyclone # SH30072.03), 1% non-essential amino acids (Hyclone #SH30238.01), 1 mM Na-pyruvate (Hyclone # SH30239.01), 10 mM HEPES (Hyclone # SH30237.01), and 1% Pen/Strep (hyclone # SV30010).
- Growth Medium 5B (BPS Cat. #79541)
- 96-well tissue culture plate or 96-well tissue culture-treated white clear-bottom assay plate
- ONE-Step™ Luciferase Assay System (BPS, Cat. #60690)
- Luminometer

Mycoplasma Testing

The cell line has been screened using the PCR-based VenorGeM Mycoplasma Detection kit (Sigma-Aldrich) to confirm the absence of *Mycoplasma* species.

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A. Dose response of Gli Reporter – NIH3T3 cells to mouse Sonic Hedgehog (mShh)

1. Harvest Gli Reporter – NIH3T3 cells from culture in Growth Medium 5B and seed cells at a density of 25,000 cells per well into white clear-bottom 96-well microplate in 100 μ l of Thaw Medium 5.
2. Incubate cells at 37°C in a CO₂ incubator for 16-20 hours.
3. The next day, the cells should reach confluency. It is critical for the cells to reach confluency in the wells before treatment.

Carefully remove the medium from the wells and avoid disrupting the cell monolayer. The cells are prone to detach at this stage. We recommend using a pipettor, not an aspirator, to remove the medium.

Add 50 μ l of threefold serial dilution of mShh in assay medium to stimulated wells.
Add 50 μ l of assay medium to the unstimulated control wells.
Add 50 μ l of assay medium to cell-free control wells (for determining background luminescence).

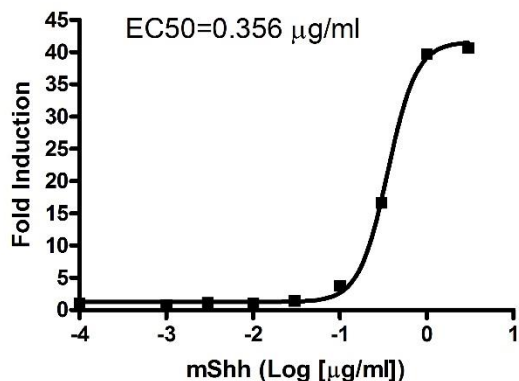
Set up each treatment in at least triplicate.

4. Incubate the plate at 37°C in a CO₂ incubator for 24 to 30 hours.
5. Perform luciferase assay using the ONE-Step™ Luciferase Assay System according to the protocol provided: Add 100 μ l of ONE-Step™ Luciferase reagent per well and rock at room temperature for ~20 minutes. Measure luminescence using a luminometer.
If using luciferase reagents from other vendors, follow the manufacturer's assay protocol.
6. Data Analysis: Subtract the average background luminescence (cell-free control wells) from the luminescence reading of all wells. The fold induction of Gli luciferase reporter expression = background-subtracted luminescence of mShh-stimulated well / average background-subtracted luminescence of unstimulated control wells.

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Figure 1. Dose Response of Gli Reporter – NIH3T3 Cells to mShh.

The results are shown as fold induction of Gli luciferase reporter expression.



B. Inhibition of mShh-induced reporter activity by inhibitors of hedgehog signaling pathway in Gli Reporter - NIH3T3 cells

1. Harvest Gli Reporter – NIH3T3 cells from culture in Growth Medium 5B and seed cells at a density of 25,000 cells per well into white clear-bottom 96-well microplate in 100 µl of Thaw Medium 5.
2. Incubate cells at 37°C in a CO₂ incubator for 16-20 hours.
3. The next day, the cells should reach confluency. It is critical for the cells to reach confluency in the wells before treatment.

Carefully remove the medium from wells and avoid disrupting the cell monolayer. The cells are prone to detach at this stage. We recommend using a pipettor, not an aspirator, to remove the medium.

Prepare stock solution of hedgehog pathway inhibitor (Cyclopamine or GDC-0449) in DMSO.

Dilute the inhibitor stock in assay medium and add 45 µl of diluted inhibitor in assay medium to the wells. The final concentration of DMSO in assay medium can be up to 0.5%.

Add 45 µl of assay medium with same concentration of DMSO without inhibitor to inhibitor control wells.

Add 45 µl of assay medium with DMSO to cell-free control wells (for determining background luminescence). (Table 1)

4. Incubate the plate at 37°C in a CO₂ incubator for 1-2 hours.

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5. Add 5 µl of diluted mShh in assay medium to stimulated wells (final [mShh] = 1 µg/ml). Add 5 µl of assay medium to the unstimulated control wells (cells without inhibitor and mShh treatment for determining the basal activity). Add 5 µl of assay medium to cell-free control wells. Set up each treatment in at least triplicate. (Table 1)

Table 1. Treatment Reference Guide

	Stimulated Wells		Unstimulated Control Wells	Cell-free Control Wells
	With inhibitor	Without inhibitor (control well)		
Step 3	45 µl diluted inhibitor in assay medium	45 µl assay medium with DMSO only	45 µl assay medium with DMSO only	45 µl assay medium with DMSO only
Step 5	5 µl mShh in assay medium (final [mShh] = 1 µg/ml)	5 µl mShh in assay medium (final [mShh] = 1 µg/ml)	5 µl assay medium	5 µl assay medium

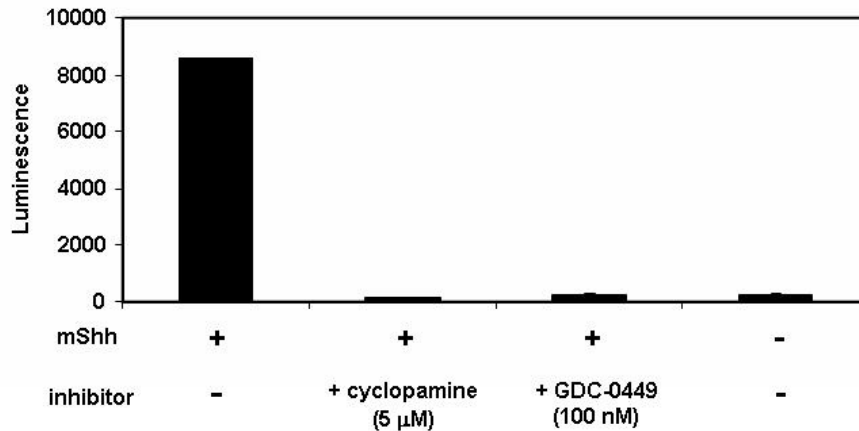
6. Incubate the plate at 37°C in a CO₂ incubator for 24-30 hours.
7. Perform luciferase assay using the ONE-Step™ Luciferase Assay System according to the protocol provided: Add 100 µl of ONE-Step™ Luciferase reagent per well and rock at room temperature for ~20 minutes. Measure luminescence using a luminometer. *If using luciferase reagents from other vendors follow the manufacturer's assay protocol.*
8. Data Analysis: Obtain the background-subtracted luminescence by subtracting the average background luminescence (cell-free control wells) from the luminescence reading of all wells.

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Figure 2. Inhibition of mShh-induced Reporter Activity by Hedgehog Pathway Inhibitors in Gli Reporter – NIH3T3 Cells

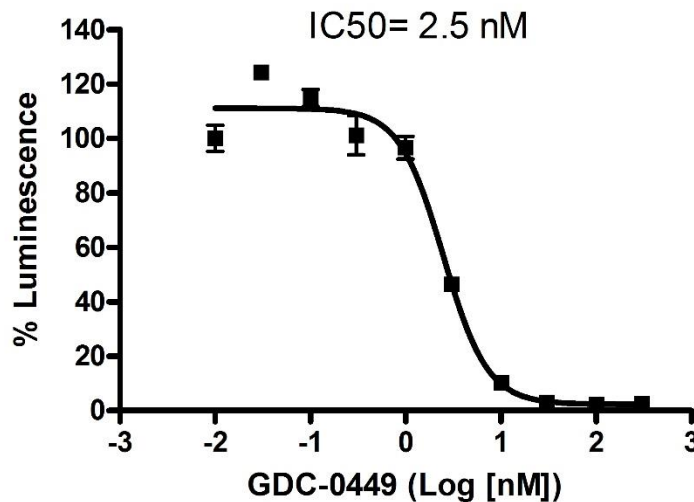
2A. Cyclopamine or GDC-0449 Inhibition of mShh-induced Gli Reporter Activity.

Raw luminescence of mShh-induced Gli reporter activity in the presence of the indicated inhibitor at the indicated concentration.



2B. GDC-0449 Inhibition Dose Response Curve

The results are shown as percentage of luminescence. The background-subtracted luminescence of cells stimulated with mShh in the absence of GDC-0449 was set at 100%.



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1. Kinzler KW *et al.* (1990) The GLI gene encodes a nuclear protein which binds specific sequences in the human genome. *Mol Cell Biol.* **10(2)**:634-642.
2. Mullor JL *et al.* (2002) Pathways and consequences: Hedgehog signaling in human disease. *Trends Cell Biol.* **12(12)**:562-569.
3. Peukert S *et al.* (2010) Small-molecule inhibitors of the hedgehog signaling pathway as cancer therapeutics. *ChemMedChem.* **5(4)**:500-512.

Related Products

<u>Product Name</u>	<u>Catalog #</u>	<u>Size</u>
Cyclopamine	27013	5 mg
Vismodegib (GDC-0449)	27010	10 mg
Wnt Signaling Pathway TCF/LEF Reporter – HEK293 Cell Line	60501	1 Vial
TCF/LEF Reporter Kit (Wnt/ β -catenin Signaling Pathway)	60500	500 Reactions
ONE-Step™ Luciferase Assay System	60690-1	10 ml
ONE-Step™ Luciferase Assay System	60690-2	100 ml

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