

Description

The NMT1 Fluorogenic Assay Kit is designed to measure NMT1 (N-myristoyltransferase 1) N-myristoylation activity for screening and profiling applications. The kit comes in a convenient 96-well format, with purified recombinant NMT1 (amino acids 110-496), NMT Peptide 1, Myristoyl – coenzyme A (MCoA), CPM (fluorescent dye), and assay buffer for 100 enzyme reactions.

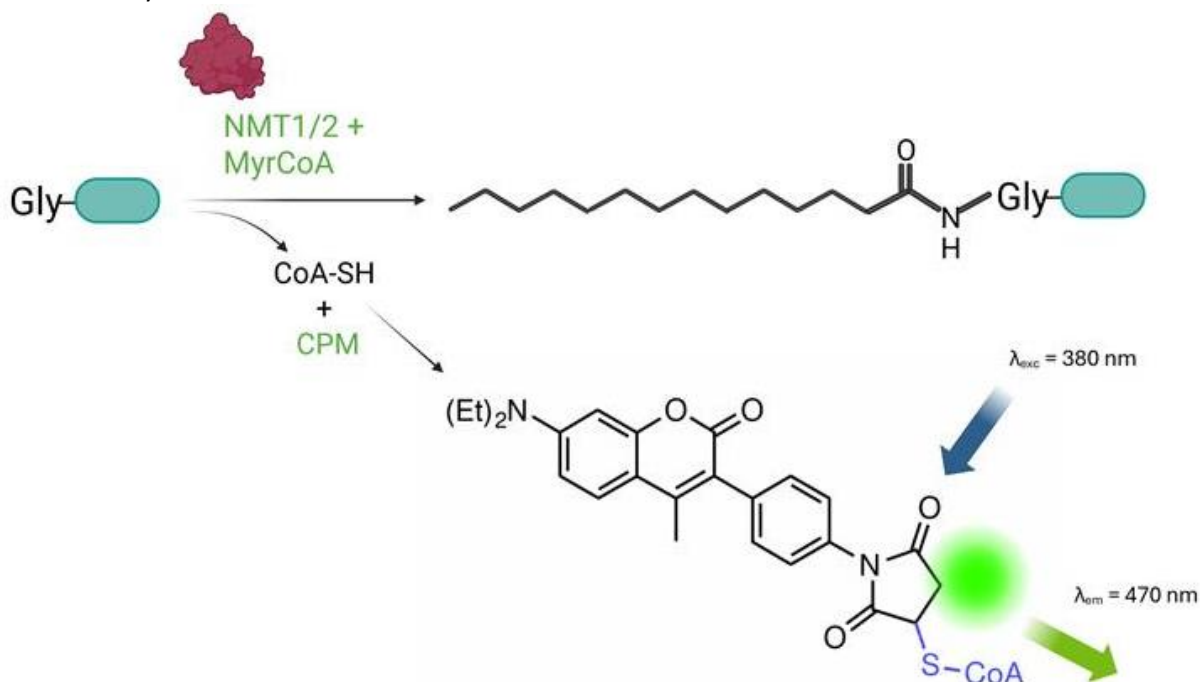


Figure 1. Mechanism of action of NMT1 Fluorogenic Assay Kit.

Background

NMT1 (N-myristoyltransferase 1), also known as Glycylpeptide N-tetradecanoyltransferase 1, is a member of the NMT family of proteins. It is a cytosolic enzyme that catalyzes the addition of a myristoyl group to glycine residues located in the N-terminus of eukaryotic, fungal and viral proteins. The addition of the myristoyl group allows for proper protein localization, and for cell survival and proliferation. It plays a role in embryonic development and cancer cell metabolism and growth. It is linked to breast, colon, and other cancer types, contributing to drug resistance. Research has found that inhibitors for NMT1 can lead to tumor growth reduction. Further studies into the role of this protein will open new treatment opportunities in cancer therapy.

Applications

Screen small molecule inhibitors or antagonists that affect activity of NMT1 in high throughput screening (HTS) applications.

Supplied Materials

Catalog #	Name	Amount	Storage
102928-KC2	NMT1, His-tag*	2 µg	-80°C
84139-KC25	NMT Peptide 1 (5 mM)	25 µl	-80°C
84140-KC5	Myristoyl – coenzyme A (MCoA) (5 mM)	5 µl	-80°C
84141-KC5	CPM (20 mM)	5 µl	-80°C
84142-KC10	1x NMT Assay Buffer	2 x 10 ml	-20°C
79685	96-well black, low binding microplate	1	Room Temp

*The concentration of the protein is lot-specific and will be indicated on the tube.

Materials Required but Not Supplied

- Adjustable micropipettor and sterile tips
- Fluorescent microplate reader capable of reading $\lambda_{exc}/\lambda_{em}$ = 380 nm/470 nm
- Orbital shaker

Storage Conditions

This assay kit will perform optimally for up to **6 months** from date of receipt when the materials are stored as directed.

Safety

This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

Contraindications

- The final concentration of DMSO in the reaction should not exceed 3%.
- Compounds that are fluorescent may interfere with the results, depending on their spectral excitation and emission properties.
- It is recommended that the compound alone is tested to determine any potential interference of the compound with the assay results.

Assay Protocol

- All samples should be run in duplicate while controls should be performed in quadruplicate.
- The assay should include “Blank”, “Positive Control” and “Test Inhibitor” conditions.
- We recommend maintaining the diluted protein on ice during use.
- For detailed information on protein handling please refer to [Protein FAQs \(bpsbioscience.com\)](https://bpsbioscience.com/protein-faqs/).
- We recommend using Zelenistat (#84233) as internal controls for the assay. If not running a dose response curve for the control inhibitor, run at 0.1X, 1X and 10X the IC₅₀ value shown in the validation data below.
- For instructions on how to prepare reagent dilutions please refer to [Serial Dilution Protocol \(bpsbioscience.com\)](https://bpsbioscience.com/serial-dilution-protocol/).

1. Thaw **1x NMT Assay Buffer** and **NMT1**. Briefly spin the tube to recover its full content.
2. Add 30 µl of 1x NMT Assay Buffer to the “Blank” wells.
3. Dilute **NMT1** to 0.67 ng/µl (30 µl/well) with 1x NMT Assay Buffer.
4. Add 30 µl of diluted NMT1 to the “Positive Control” and “Test Inhibitor” wells.
5. Prepare the **Test inhibitor** (5 µl/well): for a titration prepare serial dilutions at concentrations 10-fold higher than the desired final concentrations. The final volume of the reaction is 50 µl.

5.1 If the Test Inhibitor is water-soluble, prepare serial dilutions in 1x NMT Assay Buffer at concentrations 10-fold higher than the desired final concentrations.

For positive and negative controls, use 1x NMT Assay Buffer (Diluent Solution).

OR

5.2 If the Test inhibitor is soluble in DMSO, prepare the test inhibitor in 100% DMSO at a concentration 100-fold higher than the highest desired final concentration, then dilute the inhibitor 10-fold in 1x NMT Assay Buffer to prepare the highest concentration of the serial dilutions. The concentration of DMSO is now 10%.

Using 1x NMT Assay Buffer containing 10% DMSO to keep the concentration of DMSO constant, prepare serial dilutions of the Test Inhibitor at 10-fold the desired final concentrations.

For positive and negative controls, prepare 10% DMSO in 1x NMT Assay Buffer so that all wells contain the same amount of DMSO (Diluent Solution).

Note: The final concentration of DMSO in the assay should not exceed 1%.

6. Add 5 µl of **Test inhibitor** to each well designated “Test Inhibitor”.
7. Add 5 µl of **Diluent Solution** to the “Blank” and “Positive Control” wells.
8. Preincubate the plate for 60 minutes at Room Temperature (RT) with gentle agitation.
9. Thaw **NMT Peptide 1**, **Myristoyl – coenzyme A (MCoA)**, and **CPM**. Briefly spin the tubes to recover their full content.
10. Dilute 20-fold the **NMT Peptide 1** with 1x NMT Assay Buffer (5 µl/ well), to 250 µM.
11. Dilute **CPM** 400-fold with 1x NMT Assay Buffer (5 µl/ well), to 50 µM.
12. Dilute **MCoA** 100-fold with 1x NMT Assay Buffer (5 µl/ well), to 50 µM.

13. Prepare a **Master Mix** (15 μ l/well): N wells x (5 μ l of diluted NMT Peptide 1 + 5 μ l of diluted CPM + 5 μ l of diluted MCoA).

Note: Prepare Master Mix in excess (usually 10-20% extra) to account for pipetting losses, spills, or errors.

14. Add 15 μ l of Master Mix to all wells to initiate the reaction.

Component	Blank	Positive Control	Test Inhibitor
1x NMT Assay Buffer	30 μ l	-	-
Diluted NMT1 (0.67 ng/ μ l)	-	30 μ l	30 μ l
Test Inhibitor	-	-	5 μ l
Diluent Solution	5 μ l	5 μ l	-
Pre-incubate 60 minutes at RT			
Master Mix	15 μ l	15 μ l	15 μ l
Reaction	50 μl	50 μl	50 μl

15. Incubate at RT for 60 minutes or perform kinetic analysis.
16. Read the fluorescence intensity of the samples ($\lambda_{\text{excitation}}$ = 380 nm; $\lambda_{\text{emission}}$ = 470 nm with a bandwidth of 20 nm) in an appropriate microplate reader.
17. The “Blank” value should be subtracted from all other values.

Example Results

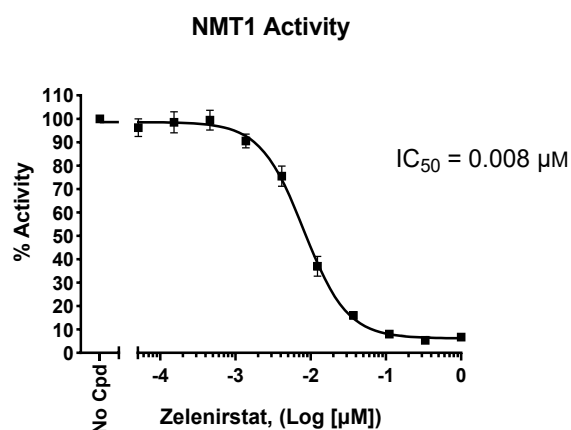


Figure 2. Inhibition of NMT1 activity by Zelenirstat.

NMT1 activity was measured in the presence of increasing concentrations of Zelenirstat. Results are expressed as percent of control activity (measured in the absence of inhibitor and set at 100%).

Data shown is representative.

Troubleshooting Guide

Visit bpsbioscience.com/assay-kits-faq for detailed troubleshooting instructions. For lot-specific information and all other questions, please visit <https://bpsbioscience.com/contact>.

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