## Description

The PARP11 Homogeneous Assay Kit is designed to measure PARP11 (poly-(ADP-ribose) polymerase 11) activity for screening and profiling applications. The PARP11 Homogeneous Assay Kit comes in a convenient 384-well AlphaLISA® format, with enough purified recombinant PARP11 (amino acids 8-338), biotinylated histone substrate, ADP-Ribose Binding Reagent 1 and assay buffer for 400 enzyme reactions.

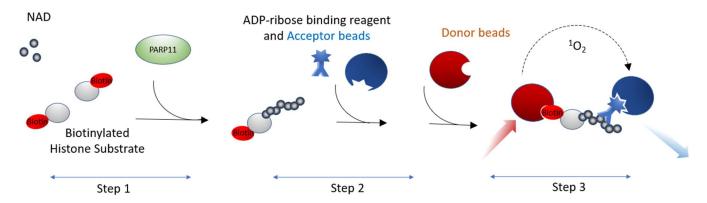


Figure 1: PARP11 Homogenous Assay Kit schematic.

A sample containing PARP11 is incubated with a biotinylated histone substrate and NAD<sup>+</sup> for one hour. This is followed by the addition of acceptor beads and ADP-Ribose Binding Reagent 1, and finally donor beads. Alphacounts are then measured. Alpha-counts are directly proportional to PARP11 activity.

## **Background:**

PARP11, also known as poly-(ADP-ribose) polymerase 11 or NAD+ ADP-ribosyltransferase 11, is part of the PARP family. ADP ribosylation, which is the addition of an ADP-ribose to a protein, is a reversible post-translational modification of proteins mostly involved in the DNA Damage Response (DDR) pathway. Mono-ADP-ribosylation (termed MARylation) is the addition of a unit of ADP-ribose. PARP11 is involved in MARylation of  $\beta$ -TrCP ( $\beta$ -transducin repeat containing protein), an E3 ligase that ubiquitinates and targets IFNAR1 (IFN $\alpha$ / $\beta$  receptor subunit 1) for degradation, restricting the potency of IFN-I (interferon type I) response to viral infections. PARP11 contributes to an immunosuppressive TME (tumor microenvironment) by downregulating IFNAR1 and decreasing the potency of CD8+ T cells to kill cancer cells. The use of inhibitors to PARP11 or engineered T cells to decrease PARP11 activity can prove beneficial in cancer therapy.

## Application(s)

Screen small molecule inhibitors for drug discovery and high throughput screening (HTS) applications.



#### **Supplied Materials**

Catalog #	Name	Amount	Storage
80511	PARP11, GST-Tag, His-Tag*	40 μg	-80°C
	5X PP-01 Assay Buffer	2 x 1 ml	-80°C
	Biotinylated Histone Substrate	500 reactions	-80°C
78311	ADP-Ribose Binding Reagent 1	10 μΙ	-80°C
	750 μM NAD <sup>+</sup>	400 μΙ	-80°C
	0.5 mM DTT	200 μΙ	-20°C
	4X Detection Buffer 1NP	2 ml	-20°C

<sup>\*</sup>The initial concentration of enzyme is lot-specific and will be indicated on the tube containing the protein.

## **Materials Required but Not Supplied**

Name	Ordering Information
AlphaLISA® Anti-Rabbit IgG Acceptor Beads	Perkin Elmer #AL104C
AlphaScreen® Streptavidin-Conjugated Donor Beads	Perkin Elmer #6760002S
Optiplate - 384	Perkin Elmer #6007290
AlphaScreen microplate reader	

#### **Storage Conditions**



This assay kit will perform optimally for up to **6 months** from date of receipt when the materials are stored as directed.

## Safety



This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

#### **Contraindications**

- The PARP11 Homogenous Assay Kit is compatible with up to 1% final DMSO concentration.
- Avoid green and blue dyes that absorb light in the AlphaScreen signal emission range ( $\lambda$ =520-620 nm), such as Trypan Blue.
- Avoid the use of potent singlet oxygen quenchers such as sodium azide (NaN<sub>3</sub>) or metal ions (Fe<sup>2+</sup>, Fe<sup>3+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup> and Ni<sup>2+</sup>).
- The presence of >1% RPMI 1640 culture medium leads to a signal reduction due to the presence of excess biotin and iron in this medium. Media like MEM, which lacks these components, does not affect AlphaScreen assays.



#### **Assay Protocol**

- All samples and controls should be tested in triplicate.
- The assay should include "Blank", "Positive Control" and "Test inhibitor".
- We recommend maintaining the diluted protein on ice during use.
- For detailed information on protein handling please refer to Protein FAQs (bpsbioscience.com).
- We recommend using Olaparib (#78318 or #27003) and Talazoparib (#78318) as internal control. If not running a dose response curve for the control inhibitor, we recommend running the control inhibitor at 0.1X, 1X and 10X the IC<sub>50</sub> value shown in the validation data below.

## Step 1:

1. Reconstitute Biotinylated Histone Substrate with 500 μl of distilled water.

Note: Diluted Biotinylated Histone Substrate can be stored into single use aliquots (minimum volume  $5 \mu l/a liquot$ ) and stored at -80°C.

- 2. Thaw 5X PP-01 Assay Buffer.
- 3. Dilute 0.5 M DTT 50-fold with 5X PP-01 Assay Buffer. This makes 5X PP-01 Assay Buffer with 10 mM DTT.

Note: Prepare only enough DTT-containing buffer as required for the assay. Store the remaining assay buffer at -20°C.

- 4. Dilute 5x PP-01 Assay Buffer with 10 mM DTT 5-fold with distilled water. This makes 1x PP-01 Assay Buffer.
- 5. Prepare the Test Inhibitor (3  $\mu$ l/well): for a titration prepare serial dilutions at concentrations 5-fold higher than the desired final concentrations. The final volume of the reaction is 15  $\mu$ l.
  - 5.1 If the Test Inhibitor is water-soluble, prepare serial dilutions in distilled water, 5-fold more concentrated than the desired final concentrations.

For the positive and negative controls, use distilled water (Diluent Solution).

#### OR

5.2 If the Test inhibitor is soluble in DMSO, prepare the test inhibitor at 100-fold the highest desired concentration in 100% DMSO, then dilute the inhibitor 20-fold in distilled water to prepare the highest concentration of the 5-fold intermediate dilutions. The concentration of DMSO is now 5%.

Prepare serial dilutions of the Test Inhibitor at 5-fold the desired final concentrations using 5% DMSO in distilled water to keep the concentration of DMSO constant.

For positive and negative controls, prepare 5% DMSO in water (vol/vol) so that all wells contain the same amount of DMSO (Diluent Solution).

Note: The final concentration of DMSO should not exceed 1%.



#### Step 2:

- 1. Make a Master Mix (7  $\mu$ l/ well): N wells × (1  $\mu$ l of reconstituted Biotinylated Histone Substrate + 1  $\mu$ l of 750  $\mu$ M NAD<sup>+</sup> + 2  $\mu$ l of 5X PP-01 Assay Buffer with 10 mM DTT + 3  $\mu$ l of distilled water).
- 2. Add 7 μl of Master Mix to each well.
- 3. Thaw PARP11 on ice. Briefly spin the tube containing enzyme to recover the full content of the tube.
- 4. Dilute PARP11 to 20 ng/μl with 1X PP-01 Assay Buffer (5 μl/well).
- 5. Add 3 μl of inhibitor solution to each well designated "Test Inhibitor".
- 6. Add 3 μl of the Diluent Solution to the "Blank" and "Positive Control" wells.
- 7. Add 5  $\mu$ l of 1x PP-01 Assay Buffer to the "Blank" wells.
- 8. Initiate the reaction by adding 5  $\mu$ l of diluted PARP11 to the wells labeled "Positive Control" and "Test Inhibitor".
- 9. Incubate at Room Temperature (RT) for 1 hour with slow agitation.

Component	Test Inhibitor	Blank	Positive Control
Master Mix	7 μΙ	7 μΙ	7 μΙ
Test Inhibitor	3 μΙ	-	-
Diluent Solution	_	3 μΙ	3 μΙ
1x PP-01 Assay Buffer	-	5 μΙ	-
Diluted PARP11 (20 ng/μl)	5 μΙ	-	5 μΙ
Total	15 μΙ	15 µl	15 μΙ



Protect your samples from direct exposure to light for step 3. Photobleaching will occur!

#### Step 3:

- 1. Dilute 4-fold the 4X Detection Buffer 1NP with distilled water. This makes 1X Detection Buffer.
- 2. Prepare a mixture containing AlphaLISA™ Anti-Rabbit Acceptor Beads diluted 250-fold and ADP-Ribose Binding Reagent 1 diluted 400-fold in 1X Detection Buffer (10 µl of mix/well).
- 3. Add 10 µl of mixture to each well.
- 4. Briefly agitate the plate.
- 5. Dilute AlphaScreen® Streptavidin-Conjugated Donor Beads 250-fold with 1x Detection Buffer (10 μl/well).



- 6. Add 10 µl per well.
- 7. Incubate for up to 1 hour at RT.

Note: First reading can be done 5-15 minutes, but longer incubation (up to 1 hour) is recommended for better readouts.

- 8. Read Alpha-counts on an AlphaScreen® microplate reader.
- 9. The "Blank" control might be important to determine the background A-screen counts in the assay. The blank value should be subtracted from all other values.

#### **Example Results**

# **PARP11 Activity**

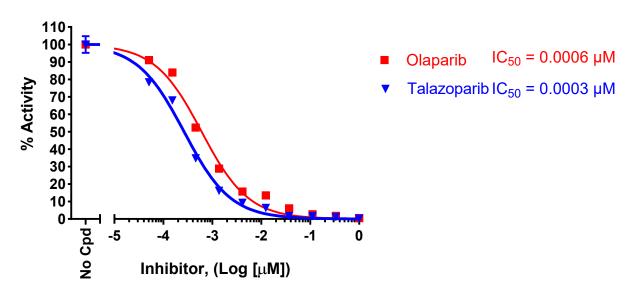


Figure 2: Inhibition of PARP11 activity by Olaparib and Talazoparib.

PARP11 was incubated with increasing concentrations of Olaparib (#78318 or #27003) and Talazoparib (#78318). Results are expressed as percent activity, in which absence of inhibitor is set to 100%.

Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at support@bpsbioscience.com.

## **Troubleshooting Guide**

Visit bpsbioscience.com/assay-kits-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com



## References

Zhang H., et al., 2022 Nat Cancer 3(7): 808-820. Guo T., et al., 2019 Nat Microbiol 4(11):1872-1884.

## **Related Products**

Products	Catalog #	Size
PARP1 Homogeneous Assay Kit	78438	384 reactions
PARP3 Homogeneous Assay Kit	78491	384 reactions
PARPtrap™ Assay Kit for PARP1	80584	96 reactions/384 reactions
PARPtrap™ Assay Kit for PARP2	78296	96 reactions/384 reactions
PARPtrap™ Combo Assay Kit for PARP1 and PARP2	78317	384 reactions
TNKS1 Homogeneous Assay Kit	78489	384 reactions

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