

**Description**

The NFAT Luciferase Reporter Lentiviruses are replication incompetent, HIV-based, VSV-G pseudotyped lentiviral particles that are ready to transduce almost all types of mammalian cells, including primary and non-dividing cells. The particles contain a firefly luciferase gene driven by the NFAT response element located upstream of the minimal TATA promoter (Figure 1) and an antibiotic selection gene (puromycin) for the selection of stable clones. After transduction, activation of the NFAT signaling pathway in the target cells can be monitored by measuring the luciferase activity.

**Application**

1. Screen for activators or inhibitors of NFAT signaling pathway in transduced target cells
2. Generate NFAT Luciferase Reporter stable cell line (puromycin resistant)

**Background**

NFAT (Nuclear factor of activated T-cells) is a family of transcription factors that has an important function in immune responses, for example by inducing the expression of various cytokines (such as IL-2-3-4 and TNF-alpha) in T cells. Members of the NFAT family have been found in many tissue types including heart, skeletal muscle and brain cells. Through their role in the immune system NFATs are involved in inflammation and these transcription factors are considered promising therapeutic targets for a variety of diseases.

NFAT is regulated by Ca<sup>2+</sup> and the Ca<sup>2+</sup>/calmodulin-dependent serine phosphatase calcineurin. NFAT proteins are phosphorylated and reside in the cytoplasm in resting cells; upon stimulation, they are dephosphorylated by calcineurin, translocate to the nucleus, and induce gene expression.

**Formulation**

The lentivirus particles were produced from HEK293T cells. They are supplied in cell culture medium containing 90% DMEM + 10% FBS.

**Titer**

Two vials (500 µl x 2) of lentivirus at a titer >10<sup>7</sup> TU/ml. The titer will vary with each lot; the exact value is provided with each shipment.

**Storage**

Lentiviruses are shipped with dry ice. For long-term storage, it is recommended to store the lentiviruses at -80°C. Avoid repeated freeze-thaw cycles. Titters can drop significantly with each freeze-thaw cycle.

**Biosafety**

The lentiviruses are produced with the SIN (self-inactivation) lentivector which ensures self-inactivation of the lentiviral construct after transduction and after integration into the genomic DNA of the target cells. None of the HIV genes (gag, pol, rev) will be expressed in the transduced cells, as they are expressed from packaging plasmids lacking the packing signal and are not present in the lentivirus particle. Although the pseudotyped lentiviruses are replication-incompetent, they require the use of a Biosafety Level 2 facility. BPS Bioscience recommends following all local federal, state, and institutional regulations and using all appropriate safety precautions.

**Materials Required but Not Supplied**

These materials are not supplied with this lentivirus but are necessary to follow the protocol described in the “Validation Data” section. Media, reagents, and luciferase assay buffers used at BPS Bioscience are all validated and optimized for use with this lentivirus and are highly recommended for best results.

Name	Ordering Information
Jurkat cells	ATCC # TIB-152
Anti-CD3 agonist antibody	<a href="#">BPS Bioscience #71274</a>
Thaw Medium 2	<a href="#">BPS Bioscience #60184</a>
Polybrene	Millipore #TR-1003-G
96-well tissue culture, clear-bottom, white plate	Corning #3610
One-Step Luciferase assay system	<a href="#">BPS Bioscience #60690</a>
Luminometer	

**Assay Protocol**

The following protocol is a general guideline for transducing Jurkat cells using the NFAT luciferase reporter lentivirus. The optimal transduction conditions (e.g. MOI, concentration of polybrene, time of assay development) should be optimized according to the cell type and the assay requirements. In most cell types, the expression of the reporter gene can be measured approximately 72 hours after transduction. For cell types with low transduction efficacy, it may be necessary to select the cells stably expressing the reporter gene with puromycin prior to carrying out the reporter assays.

1. Day 1: Harvest the Jurkat cells by centrifugation and resuspend the cells in fresh Thaw Medium 2. Dilute the cells to a density of  $2 \times 10^5$  /ml in medium. Mix 500  $\mu$ l of the Jurkat cells and 400  $\mu$ l of NFAT luciferase reporter lentivirus in a 1.5-ml Eppendorf tube (at a MOI >10).

Add polybrene to a final concentration of 8  $\mu$ g/ml. Gently mix and incubate the virus with the Jurkat cells for 20 minutes at room temperature in the tissue culture hood.

Centrifuge the virus/cell mixture for 30 minutes at 800 x g at 32°C. Remove the virus-containing medium and resuspend the cell pellet in 2 ml of fresh Thaw Medium 2. Transfer the cells into one well of a 6-well plate. Incubate the plate at 37°C with 5% CO<sub>2</sub> for 48-66 hours. The transduced Jurkat cells are ready for assay development on day 4.

2. Day 3: Coat a cell culture-treated, clear bottom, white 96-well plate with anti-CD3 antibody diluted in PBS (Phosphate Buffer Saline) overnight. Leave a few non-coated wells to serve as negative controls.
3. Day 4: wash all wells of the coated plate three times with PBS. The plate is ready to use.
4. Day 4: Harvest the transduced Jurkat cells (from day 1) and resuspend the cells into 900  $\mu$ l of fresh Thaw Medium 2. Add 100  $\mu$ l of cells to each well of the CD3 antibody-coated 96-well plate.
5. Incubate at 37°C with 5% CO<sub>2</sub> for 5-6 hours.

- Prepare the ONE-Step™ Luciferase reagent per recommended protocol (100 µl/well). Add 100 µl of ONE-Step™ Luciferase Assay reagent per well. Incubate at room temperature for ~15 to 30 minutes and measure luminescence using a luminometer.

**Validation Data**

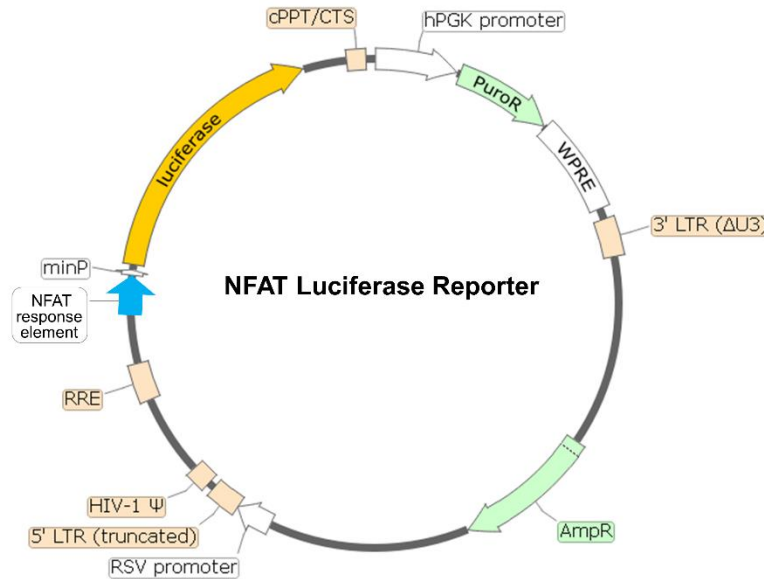


Figure 1: Schematic of the lenti-vector used to generate the NFAT luciferase reporter lentivirus

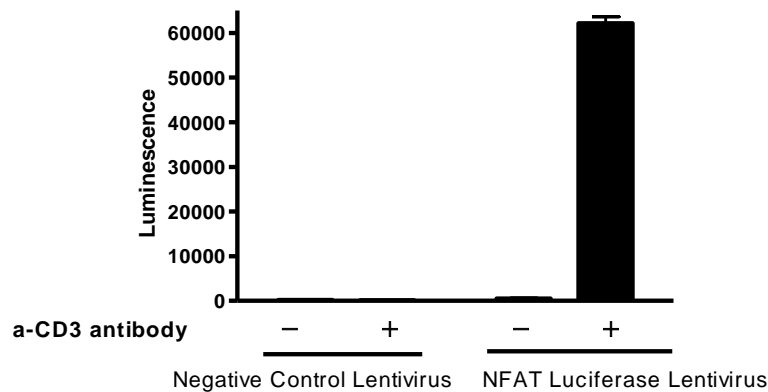


Figure 2. NFAT luciferase reporter activity stimulated by anti-CD3 agonist antibody in Jurkat cells. Appropriate 20,000 Jurkat cells were transduced with 300,000 TU NFAT luciferase reporter lentivirus. After 66 hours of transduction, medium was changed to Thaw medium 2. Cells were stimulated with anti-CD3 agonist antibody (precoated on a 96-well plate) for ~6 hours. The noncoated wells and the negative control lentivirus were performed in parallel as controls. The results are shown as the raw luminescence reading.

**Notes**

1. To generate the NFAT luciferase reporter stable cell line, remove the growth medium 48 hours after transduction and replace it with fresh growth medium containing the appropriate amount of puromycin for antibiotic selection of transduced cells.
2. The following Lentivirus Reporter Controls are available from BPS Bioscience to meet your experimental needs:
  - a) Negative Control Luciferase Lentivirus (BPS Bioscience #79578): Ready-to-transduce lentiviral particles expressing firefly luciferase under the control of a minimal promoter. The negative control is important to establish the specificity of any treatments and to determine the background reporter activity.
  - b) Renilla Luciferase Lentivirus (BPS Bioscience #79565): Ready-to-transduce lentiviral particles expressing Renilla luciferase under the CMV promoter. The RLuc lentivirus can serve as an internal control to overcome sample-to-sample variability when performing dual-luciferase reporter assays.
  - c) Firefly Luciferase Lentivirus (BPS Bioscience #79692-G, #79692-H, #79692-P): Ready-to-transduce lentiviral particles expressing firefly luciferase under the CMV promoter. It serves as a positive control for transduction optimization studies.

**Reading Luminescence**

Luminescence is the emission of light resulting from a chemical reaction. The detection of luminescence requires no wavelength selection because the method used is emission photometry and not emission spectrophotometry.

To properly read luminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader: use the “hole” position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay.

**References**

1. Clipstone NA, Crabtree GR. *Nature*. 1992 Jun 25;357(6380):695-7.
2. Lyakh, L., *et al.* *Mol Cell Biol*. 1997 May;17(5):2475-84.

**Troubleshooting Guide**

Visit [bpsbioscience.com/lentivirus-faq](https://bpsbioscience.com/lentivirus-faq) for detailed troubleshooting instructions. For all further questions, please email [support@bpsbioscience.com](mailto:support@bpsbioscience.com).

**Related Products**

<i>Products</i>	<i>Catalog #</i>	<i>Size</i>
NFAT Reporter (Luciferase) – THP-1 Cell Line	78320	500 µl x 2
NFAT eGFP Reporter Lentivirus	79922	500 µl x 2