

# Data Sheet FGFR3 (L496V) Assay Kit Catalog #79818 96 Reactions

**DESCRIPTION:** The *FGFR3 (L496V) Assay Kit* is designed to measure FGFR3 (L496V) activity for screening and profiling applications using Kinase-Glo<sup>®</sup> MAX as a detection reagent. The *FGFR3 (L496V) Assay Kit* comes in a convenient 96-well format, with enough purified recombinant FGFR3 (L496V), Protein Tyrosine Kinase Substrate (Poly-Glu,Tyr 4:1), ATP, and kinase assay buffer for 96 enzyme reactions.

#### **COMPONENTS:**

Catalog #	Reagent	Amount	Stora	ge
100142	FGFR3 (L496V), His-Tag	20 µg	-80°C	Avoid
79334	5x Kinase assay buffer 1	1.5 ml	-20°C	multiple
79686	ATP (500 μM)	100 µl	-20°C	freeze/
40217	Protein Tyrosine Kinase Substrate (Poly-Glu,Tyr 4:1) (10 mg/ml)	100 µl	-20°C	thaw cycles!
79696	96-well plate, white	1	RT	

### MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:

Kinase-Glo MAX (Promega #V6071) Dithiothreitol (DTT, 0.5 M; optional) Microplate reader capable of reading luminescence Adjustable micropipettor and sterile tips 30°C incubator

**APPLICATIONS:** Useful for studying enzyme kinetics and screening small molecular inhibitors for drug discovery and HTS applications.

**STABILITY:** Up to 6 months when stored as recommended.

#### **REFERENCE:**

1. Cappellen, D., *et al.* (1999) "Frequent activating mutations of FGFR3 in human bladder and cervix carcinomas." *Nature Genetics* **23(1):** 18.

2. Billerey, C., *et al.* (2001) "Frequent FGFR3 mutations in papillary non-invasive bladder (pTa) tumors." *Amer. J. Pathology* **158(6):** 1955-1959.

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## ASSAY PROTOCOL:

All samples and controls should be tested in duplicate.

 Thaw 5x Kinase assay buffer 1, ATP (500 μM), and Protein Tyrosine Kinase Substrate (Poly-Glu,Tyr 4:1).

(Optional: If desired, add 30 µl of 0.5 M DTT to 5x Kinase assay buffer 1).

 Prepare the master mixture (25 μl per well): N wells x (10 μl 5x Kinase assay buffer 1 + 1 μl ATP (500 μM) + 1 μl Protein Tyrosine Kinase Substrate (Poly-Glu,Tyr 4:1) + 13 μl distilled water). Add 25 μl to every well.

	Positive Control	Test Inhibitor	Blank
5x Kinase assay buffer 1	10 µl	10 µl	10 µl
ΑΤΡ (500 μΜ)	1 µl	1 µl	1 µl
Poly-Glu,Tyr(10 mg/ml)	1 µl	1 µl	1 µl
Water	13 µl	13 µl	13 µl
Test Inhibitor	-	5 µl	_
Inhibitor buffer	5 µl	-	5 µl
1x Kinase buffer 1	-	-	20 µl
FGFR3 (L496V), His-tag (10 ng/µl)	20 µl	20 µl	_
Total	50 µl	50 µl	50 µl

3) Add 5 μl of Inhibitor solution of each well labeled as "Test Inhibitor." For the "Positive Control" and "Blank," add 5 μl of Inhibitor buffer (the same solution without the inhibitor. For compounds dissolved in DMSO, this is usually 10% DMSO in water).

Note: Final DMSO concentration must be  $\leq 1\%$ . Higher DMSO levels can significantly decrease the enzyme activity. For example, to test an inhibitor dissolved in 100% DMSO at 10  $\mu$ M, dilute 1 mM inhibitor with water to make a 100  $\mu$ M inhibitor in 10% DMSO(aq). Then, add 5  $\mu$ l of the 100  $\mu$ M solution into the 50  $\mu$ l assay to make a 1% DMSO concentration in the final reaction mixture.

- Prepare 3 ml of 1x Kinase assay buffer 1 by mixing 600 μl of 5x Kinase assay buffer 1 with 2400 μl water. 3 ml of 1x Kinase assay buffer 1 is sufficient for 100 reactions.
- 5) To the wells designated as "Blank," add 20 µl of **1x Kinase assay buffer 1**.

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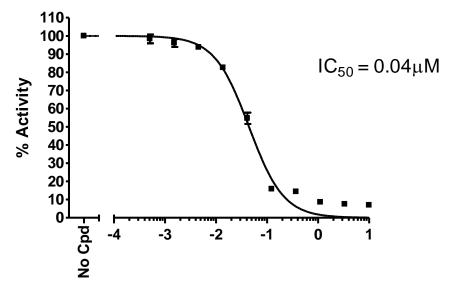
- 6) Thaw FGFR3 (L496V), His-Tag on ice. Upon first thaw, briefly spin tube containing enzyme to recover full content of the tube. Calculate the amount of FGFR3 (L496V), His-Tag required for the assay and dilute enzyme to 10 ng/µl with 1x Kinase assay buffer 1. Store remaining undiluted enzyme in aliquots at -80°C. <u>Note</u>: FGFR3 (L496V), His-Tag is sensitive to freeze/thaw cycles. Avoid multiple freeze/thaw cycles. Do not re-use thawed aliquots or diluted enzyme.
- Initiate reaction by adding 20 µl of diluted FGFR3 (L496V), His-Tag to the wells designated "Positive Control" and "Test Inhibitor Control." Incubate at 30°C for 45 minutes.
- 8) Thaw Kinase-Glo Max reagent.
- After the 45 minute reaction, add 50 µl of Kinase-Glo Max reagent to each well. Cover plate with aluminum foil and incubate the plate at room temperature for 15 minutes.
- 10) Measure luminescence using the microplate reader. Value of "Blank" reading should be subtracted from all other measurements.

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**Example of Assay Results:** 





## Erdafitinib, (Log [µM])

Inhibition of FGFR3 (L496V), His-Tag by Erdafitinib, measured using the FGFR3 (L496V) assay kit (BPS Bioscience #79818). Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at <u>support@bpsbioscience.com</u>

#### **RELATED PRODUCTS:**

Product Name	Catalog #	<u>Size</u>
FGFR3 (L496V), His-Tag	100142	10 µg
FGFR3 (V443M), His-Tag	100140	10 µg
FGFR3 (V443L), His-Tag	100138	10 µg
FGFR3 (CD333), GST-tag	40212	10 µg
5x Kinase assay buffer 1	79334	10 ml
ΑΤΡ (500 μΜ)	79686	200 µl
Protein Tyrosine Kinase Substrate		
(poly-Glu,Tyr 4:1)	40217	1 mg
FGFR1(FLT2), GST-tag	40210	10 µg
FGFR1 (V561M), GST-tag	40209	10 µg
FGFR2, GST-tag	40211	10 µg
FGFR4, GST-tag	40213	10 µg

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