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## **Data Sheet**

### **CD4<sup>+</sup> T cells, Negatively Selected (Human) Catalog # 79752**

#### **Description**

Cryopreserved vial (10 x 10<sup>6</sup> cells) of CD4<sup>+</sup> T cells that were negatively selected from freshly isolated primary human peripheral blood mononuclear cells (PBMCs). The PBMCs came from a healthy donor and were isolated from whole blood or leukapheresis samples using a Ficoll gradient. Magnetic antibodies to monocytes, granulocytes, CD8<sup>+</sup> T cells, gamma/delta T cells and other immune subsets present in PBMCs were then used to purify untouched CD4<sup>+</sup> T cells via immunomagnetic separation. Before and after CD4<sup>+</sup> T cell isolation, the cells were stained to evaluate purity and viability by flow cytometry. Cells were cryopreserved in CryoStor CS10 cryopreservation medium (Stemcell #07930) at a controlled rate.

#### **Source**

Normal human PBMC from Leukapheresis Sample

#### **Stability and Storage**

Store cells at -135°C or colder. Thawed cells should be used immediately for downstream applications. Because these are primary cells, we do not recommend maintaining these cells in culture for long periods of time.

#### **Characterization Criteria**

Cell count, viability (trypan blue exclusion and FACS with impermeable DNA binding dye), and surface expression of CD3 and CD4

#### **Medium**

**TCellIM™ (BPS Bioscience #78753):** Contains Iscove's MDM, Heat inactivated Fetal Bovine Serum (HiFBS), 1% penicillin/streptomycin, 2-Mercaptoethanol, insulin, and other supplements.

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## Handling Directions

First, pre-warm Immuno Culture Medium to 37°C. It is important to work quickly in the following steps to ensure high cell viability and recovery. Quickly thaw cells in a 37°C water bath with constant but gentle agitation. Clean the outside of the vial with 70% ethanol and immediately transfer the entire contents to a 50 ml tube. Slowly add 10 ml of pre-warmed medium while gently swirling the tube to mix. Centrifuge the cell suspension at 300 x g for 15 minutes at room temperature. Carefully remove the supernatant with a pipette without disturbing the pellet. Gently resuspend the cell pellet by flicking the tube, then add the desired volume of warm medium, and mix. NOTE: Up to 30% cell loss can be expected during washing steps. Cells are now ready for use in downstream applications.

## Donor Screening

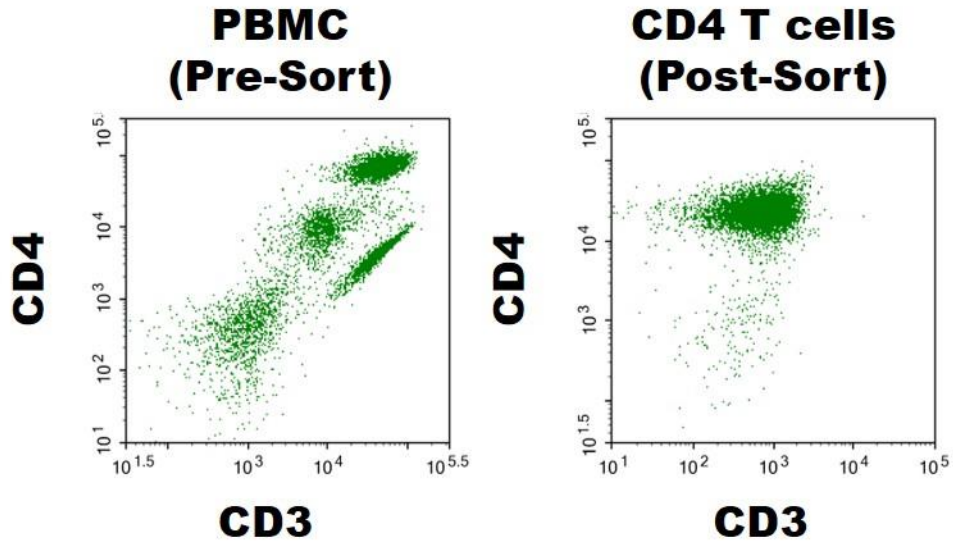
Donors have been screened and determined negative for:

- Hepatitis B (anti-HBc EIA, HBsAg EIA)
- Hepatitis C (anti-HCV EIA)
- Human Immunodeficiency Virus (HIV-1/HIV-2 plus O)
- Human T-Lymphotropic Virus (HTLV-I/II)
- HIV-1/HCV/HBV
- West Nile Virus
- Trypanasoma cruzi

**Note:** *Testing cannot guarantee that any sample is completely virus-free. These cells should be treated as potentially infectious and appropriate biological safety level 2 precautions should be used.*

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**Representative Flow Cytometry Analysis**



**Related Products**

<u>Product</u>	<u>Cat. #</u>	<u>Size</u>
Human CD8+ T cells	79753	10 x 10 <sup>6</sup> cells
Normal Human PBMC	79059	30 x 10 <sup>6</sup> cells
Thaw Medium 2	60184	100 ml
Human IL-2	90184-A	10 µg

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