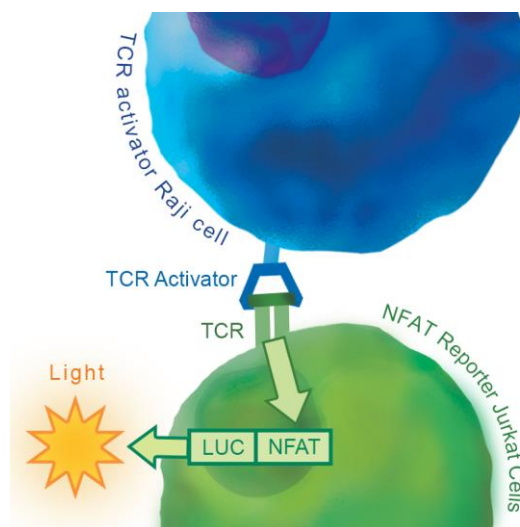


**Description**

Recombinant Raji cell line stably expressing an engineered, membrane-bound T cell receptor (TCR) activator. The cell line has been functionally validated in a co-culture assay for activation of the NFAT Reporter Jurkat cells.

**Background**

The T Cell Receptor (TCR) is a heterodimer protein found at the surface of T cells, which is stimulated upon binding to Major Histocompatibility Complex (MHC)/peptide complexes and triggers a signaling cascade that leads to the activation of transcription factors involved in the upregulation and secretion of cytokines, T cell proliferation, and cell differentiation into effector and memory cells.

The Raji line was established from a Burkitt's lymphoma patient. Raji cells constitutively express B cell antigens CD19, CD20, and CD22, and offer a physiologically relevant platform to evaluate cancer-directed immunotherapies such as Chimeric Antigen Receptor (CAR) T-cells. Raji Recombinant Cell Lines are biologically relevant targets for CAR-T or NK cells targeting CD19, CD20, or CD22.

When co-cultured with T cells, the engineered TCR activator cells stimulate the TCR on T cells, leading to activation of downstream signaling pathways and biological responses such as proliferation.

**Application**

- Activate T cells in co-culture assays
- Optimize co-culture assays with Luciferase Reporter T cells

**Materials Provided**

Components	Format
2 vials of frozen cells	Each vial contains $1 \times 10^6$ cells in 1 ml of cell freezing medium (BPS Bioscience #79796)

**Parental Cell Line**

Raji, a human B lymphoblastoid cell line derived from a patient with Burkitt's lymphoma, suspension

**Mycoplasma Testing**

The cell line has been screened to confirm the absence of Mycoplasma species.

**Materials Required but Not Supplied**

These materials are not supplied with the cell line but are necessary for cell culture and cellular assays. BPS Bioscience's reagents are validated and optimized for use with this cell line and are highly recommended for best results. Media components are provided in the Media Formulations section below.

*Media Required for Cell Culture*

Name	Ordering Information
Thaw Medium 2	<a href="#">BPS Bioscience #60184</a>
Growth Medium 2D	<a href="#">BPS Bioscience #79639</a>

**Storage Conditions**

Cells are shipped in dry ice and should immediately be thawed or stored in liquid nitrogen upon receipt. Do not use a -80°C freezer for long term storage. Contact technical support at [support@bpsbioscience.com](mailto:support@bpsbioscience.com) if the cells are not frozen in dry ice upon arrival.

**Media Formulations**

For best results, it is *highly recommended* to use these validated and optimized media from BPS Bioscience. Other preparations or formulations of media may result in suboptimal performance.



Note: Thaw Media do *not* contain selective antibiotics. However, Growth Media *do* contain selective antibiotics, which are used for maintaining the presence of the transfected gene(s) over passages.

Cells should be grown at 37°C with 5% CO<sub>2</sub>. BPS Bioscience's cell lines are stable for at least 15 passages when grown under proper conditions.

*Media Required for Cell Culture*

*Thaw Medium 2 (BPS Bioscience #60184):*

RPMI1640 medium supplemented with 10% FBS, and 1% Penicillin/Streptomycin.

*Growth Medium 2D (BPS Bioscience #79639):*

RPMI1640 medium supplemented with 10% FBS, and 1% Penicillin/Streptomycin plus 200 µg/ml of Hygromycin B.

**Cell Culture Protocol***Cell Thawing*

1. Swirl the vial of frozen cells for approximately 60 seconds in a 37°C water bath. As soon as the cells are thawed (it may be slightly faster or slower than 60 seconds), quickly transfer the entire contents of the vial to a tube containing 10 ml of pre-warmed Thaw Medium 2 (**no Hygromycin**).  
**Leaving the cells in the water bath at 37°C for too long will result in rapid loss of viability.**
2. Immediately spin down the cells at 300 x g for 5 minutes, remove the medium and resuspend the cells in 5 ml of pre-warmed Thaw Medium 2 (**no Hygromycin**).
3. Transfer the resuspended cells to a T25 flask and incubate at 37°C in a 5% CO<sub>2</sub> incubator.
4. After 24 hours of culture, check for cell viability. For a T25 flask, add 3-4 ml of Thaw Medium 2 (**no Hygromycin**), and continue growing in a 5% CO<sub>2</sub> incubator at 37°C until the cells are ready to passage.

- Cells should be passaged before they reach a density of  $2 \times 10^6$  cells/ml. At first passage and subsequent passages, use Growth Medium 2D (**contains Hygromycin**).

#### Cell Passage

Dilute the cell suspension into new culture vessels before they reach a density of  $2 \times 10^6$  cells/ml, at no less than  $0.2 \times 10^6$  cells/ml of Growth Medium 2D (**contains Hygromycin**). The sub-cultivation ratio should maintain the cells between  $0.2 \times 10^6$  cells/ml and  $2 \times 10^6$  cells/ml.

#### Cell Freezing

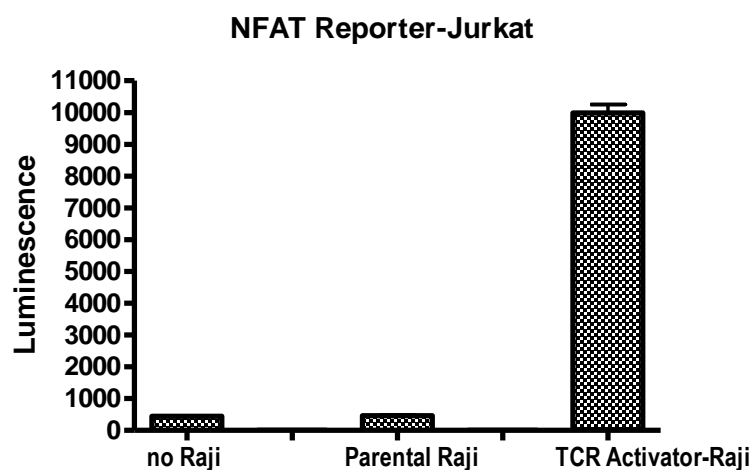
- Spin down the cells at  $300 \times g$  for 5 minutes, remove the medium and resuspend the cell pellet in  $4^\circ\text{C}$  Freezing Medium (BPS Bioscience #79796, or 10% DMSO + 90% FBS) at a density of  $\sim 2 \times 10^6$  cells/ml.
- Dispense 1 ml of cell aliquots into cryogenic vials. Place the vials in an insulated container for slow cooling and store at  $-80^\circ\text{C}$  overnight.
- Transfer the vials to liquid nitrogen the next day for storage.



Note: It is recommended to expand the cells and freeze at least 10 vials at an early passage for future use.

#### A. Validation Data

The functionality of the cell line was validated using a luciferase reporter cell-based assay. In this assay, Jurkat T cells expressing the luciferase reporter under the control of NFAT response elements are co-cultivated with TCR activator Raji cells. TCR complexes on the Jurkat cells are activated by the TCR activator expressed in Raji cells, resulting in induction of the NFAT-dependent luciferase reporter.



*Figure 1: Co-culture of TCR activator Raji cells with NFAT Luciferase Reporter Jurkat cells.*

TCR activator Raji cells or parental Raji cells were seeded in a white, clear-bottom 96-well plate. The following day, NFAT Luciferase Reporter Jurkat cells were added to the Raji cells. A negative control consisting of reporter cells without Raji cells was also performed. Induction of luciferase by the TCR activator was quantified using ONE-Step™ Luciferase System (BPS Bioscience 60690). Results are shown as raw luminescence signal.

**License Disclosure**

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**Troubleshooting Guide**

Visit [bpsbioscience.com/cell-line-faq](https://bpsbioscience.com/cell-line-faq) for detailed troubleshooting instructions. For all further questions, please email [support@bpsbioscience.com](mailto:support@bpsbioscience.com).

**Related Products**

<i>Products</i>	<i>Catalog #</i>	<i>Size</i>
TCR activator/PD-L1 CHO cell line	60536	2 vials
NFAT Reporter Jurkat cell line	60621	2 vials
TCR Knockout Jurkat Cell Line	78539	2 vials
TCR/B2M Knockout NFAT Luciferase Reporter Jurkat Cell Line	78364	2 vials
TCR Activator Lentivirus (CMV Promoter/Puromycin) or (EF1a Promoter/Puromycin) or (EF1a Promoter/Hygromycin)	79894	500 µl x 2
Firefly Luciferase Raji Cell Line	78622	2 vials
Cas9-Expressing Raji Cell Line	78156	2 vials