

Description

V γ 9V δ 2 TCR Lentivirus are replication incompetent, HIV-based, VSV-G-pseudotyped lentiviral particles ready to transduce nearly all types of mammalian cells, including primary and non-dividing cells. These viruses transduce a TCR (T Cell Receptor) which belongs to the V γ 9V δ 2 subtype of $\gamma\delta$ TCRs. The TCR γ chain and δ chain are linked by P2A. The lentiviruses also transduce a puromycin selection marker (Figure 1).

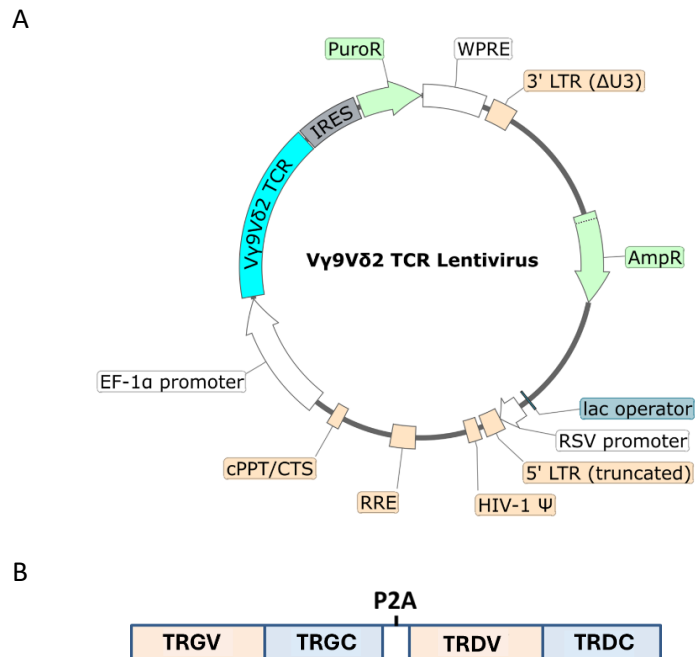


Figure 1. (A) Schematic of the lenti-vector used to generate the V γ 9V δ 2 TCR Lentivirus and (B) diagram of the construct, showing the components of the V γ 9V δ 2 TCR.

TRGV and TRGC correspond to the TCR γ chain variable and constant regions, respectively, whereas TRDV and TRDC correspond to the TCR δ chain variable and constant regions.

Background

$\gamma\delta$ TCRs (T cell receptors), $\alpha\beta$ TCRs, and antibodies, result from gene rearrangements and offer the immune system the possibility to recognize several different types of antigens. $\gamma\delta$ TCRs recognize antigens in a similar way to antibodies, being able to recognize full protein antigens and being independent on antigen binding to the MHC (major histocompatibility complex). Examples of such antigens include pyrophosphomonoesters from *Mycobacterium tuberculosis* and *E. Coli*, or glycoprotein I from *Herpes simplex*. $\gamma\delta$ TCRs are cell type specific, with V γ 9V δ 2 being present in T cells and corresponding to about 5% of the T cell population in blood. V γ 9V δ 2 T cells are involved mostly in immune responses to pathogens and long-term modulation of inflammation, and can recognize non-peptide phospho-antigens, alkylamines and synthetic aminobisphosphonates. V γ 9V δ 2 T cells are being studied for the treatment of solid tumors and hematological disorders and are becoming a highly promising cancer therapy. Further studies on how best to utilize V γ 9V δ 2 T cells, and methods to enhance their presence, will open new therapeutic avenues for cancer and infections.

Application

- Use to transduce cells with the goal of screening V γ 9V δ 2 TCR agonist antibodies.
- Use to transduce cells to be used as a positive control for V γ 9V δ 2 TCR evaluation and optimize experimental conditions.
- Generate V γ 9V δ 2 TCR expressing cell pools or stable cell lines, following puromycin selection.

Formulation

The lentiviruses were produced in HEK293T cells in medium containing 90% DMEM + 10% FBS. Virus particles can be packaged in custom formulations by special request, for an additional fee.

Size and Titer

$\geq 2 \times 10^7$ TU/ml. The titer will vary with each lot; the exact value is provided with each shipment.

Storage

Lentiviruses are shipped with dry ice. For long-term storage, it is recommended to store the virus at -80°C for up to 12 months from date of receipt. Avoid repeated freeze-thaw cycles. Titers can drop significantly with each freeze-thaw cycle.

Biosafety

The lentiviruses are produced with a SIN (self-inactivation) lentivector which ensures self-inactivation of the lentiviral construct after transduction and integration into the genomic DNA of the target cells. None of the HIV genes (*gag*, *pol*, *rev*) will be expressed in the transduced cells, as they are expressed from packaging plasmids lacking the packing signal. Although the pseudotyped lentiviruses are replication-incompetent, they require the use of a Biosafety Level 2 facility. BPS Bioscience recommends following all local federal, state, and institutional regulations and using all appropriate safety precautions.

Materials Required but Not Supplied

These materials are not supplied with this lentivirus but are necessary to follow the designed protocol. BPS Bioscience media, reagents, and luciferase assay systems are all validated and optimized for use with this lentivirus and are highly recommended for best results.

Name	Ordering Information
Thaw Medium 2	BPS Bioscience #60184
Growth Medium 2B	BPS Bioscience #79530
Growth Medium 2F	BPS Bioscience #79669
TCR Knockout NFAT-Luciferase Reporter Jurkat Cell Line	BPS Bioscience #78556
Anti-CD3 Agonist Antibody	BPS Bioscience #71274
TCR V δ 2 Antibody, anti-human, PE, REAfinity™	Miltenyi Biotec #130-111-127
TCR V delta 2 Monoclonal Antibody (15D)	Life Technologies #TCR1732
TCR V delta 1 Monoclonal Antibody (TS8.2)	Life Technologies #TCR1730
ONE-Step™ Luciferase Assay System	BPS Bioscience #60690
Lenti-Fuse Polybrene Viral Transduction Enhancer	BPS Bioscience #78939

Media Formulations

For best results, the use of BPS Bioscience validated and optimized media is *highly recommended*. Other preparations or formulations of media may result in suboptimal performance.

Media Required for Cell Culture

Growth Medium 2B (BPS Bioscience #79530):

RPMI 1640 medium supplemented with 10% FBS, 1% Penicillin/Streptomycin plus 1 mg/ml Geneticin.

Growth Medium 2F (BPS Bioscience #79669):

RPMI1640 medium supplemented with 10% FBS, 1% Penicillin/Streptomycin plus 0.5 µg/ml of Puromycin and 1 mg/ml Geneticin.

Media Used in Functional Cellular Assay

Thaw Medium 2 (BPS Bioscience #60184):

RPMI 1640 medium supplemented with 10% FBS, and 1% Penicillin/Streptomycin.

Assay Protocol

- The following protocol was used to transduce a Jurkat cell line. The transduction conditions (e.g. MOI, concentration of polybrene, time of assay development) should be optimized according to the cell type and the assay requirements. In most cell types, the expression of the reporter gene can be measured approximately 48-72 hours after transduction. For cell types with low transduction efficacy, it may be necessary to select the cells stably expressing the reporter gene with puromycin prior to carrying out the reporter assays.
- The assay should include “Negative Control” (uncoated wells) and “Testing” wells.
- 1 ml of Jurkat cells at 2×10^5 cells/ml is enough for 12 wells of a 96-well plate.

Day 1:

1. Harvest TCR Knockout NFAT-Luciferase Reporter Jurkat cells from Growth Medium 2B by centrifugation and resuspend the cells in fresh Thaw Medium 2 and count.
2. Dilute cells to a density of 2×10^5 cells/ml in Thaw Medium 2.
3. Mix 1 ml of the Jurkat cells with the appropriate amount of Vγ9Vδ2 TCR Lentivirus in a 1.5-ml Eppendorf tube.

Note: The MOI may need to be optimized, we recommend a starting MOI of 5.

4. Add Lenti-Fuse™ Polybrene Viral Transduction Enhancer to a final concentration of 8 µg/ml.
5. Gently mix and incubate the virus with the Jurkat cells for 20 minutes at Room Temperature (RT) in a tissue culture hood.
6. Centrifuge the virus/cell mixture for 30-120 minutes at $800 \times g$ and 32°C (spinoculation).
7. Add the cells/virus mix from the spinoculation step to one well of a 6-well plate.
8. Add an additional 1.5 ml of Thaw Medium 2 to the well.

Note: It is not necessary to remove the virus.

9. Incubate the cells at 37°C with 5% CO₂ for 48-66 hours.

Day 3:

1. The expression of TCR can be analyzed by flow cytometry. The transduced Jurkat cells are ready for assay development on day 3 or 4.

Note: If the transduction efficiency is low, it may be necessary to initiate cell selection with puromycin on day 3.

2. Coat a cell culture-treated, clear bottom, white 96-well plate with 100 μ l of control or agonist antibodies at 10 μ g/ml in PBS (Phosphate Buffer Saline) at 4°C overnight. Leave a few non-coated wells as “Negative Control”.

Day 4:

1. Wash all wells of the coated plate three times with PBS. The plate is ready to use.
2. Harvest the transduced Jurkat cells and resuspend cells into 1.2 ml of fresh Thaw Medium 2.
3. Add 100 μ l of cells to each well of the antibody-coated 96-well plate.
4. Incubate the plate at 37°C with 5% CO₂ for 5-6 hours.
5. Add 100 μ l of ONE-Step™ Luciferase Assay reagent per well.
6. Incubate at RT for ~15 to 30 minutes.
7. Measure luminescence using a luminometer.

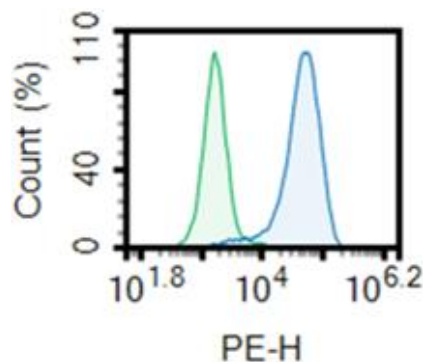
Validation Data

Figure 2. Analysis of the expression of V γ 9V δ 2 TCR by flow cytometry in TCR Knockout NFAT-Luciferase Reporter Jurkat cells transduced with the V γ 9V δ 2 TCR Lentivirus.

Approximately 100,000 TCR Knockout NFAT-Luciferase Reporter Jurkat cells (BPS Bioscience #78556) were transduced with V γ 9V δ 2 TCR Lentivirus by spinoculation at a MOI of 10. Transduced cells were selected with Growth Medium 2F and stained with TCR V δ 2 Antibody, anti-human, PE, REAfinity™ (Miltenyi Biotech #130-111-127) and analyzed by flow cytometry. The y axis represents the % of cells. The x axis indicates fluorophore intensity. Green, untransduced cells; Blue, V γ 9V δ 2 TCR transduced cells.

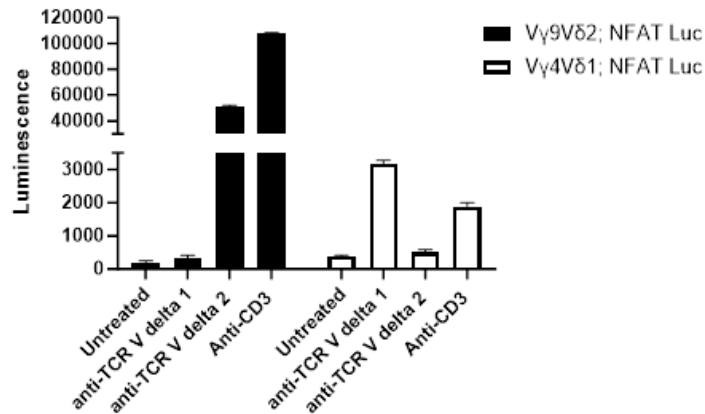


Figure 3. NFAT luciferase reporter activity in TCR Knockout NFAT-Luciferase Reporter Jurkat cells transduced with V γ 9V δ 2 TCR Lentivirus and V γ 4V δ 1 TCR Lentivirus and stimulated with agonist antibodies.

TCR Knockout NFAT-Luciferase Reporter Jurkat cells (BPS Bioscience #78556) were transduced with V γ 9V δ 2 TCR Lentivirus (BPS Bioscience #78985) or V γ 4V δ 1 TCR Lentivirus (BPS Bioscience #78986). Transduced cells were selected with Growth Medium 2F and stimulated with plate-coated TCR V delta 1 Monoclonal Antibody (TS8.2) (Life Technologies #TCR1730), TCR V delta 2 Monoclonal Antibody (15D) (Life Technologies #TCR1732), and Anti-CD3 Agonist Antibody (BPS Bioscience #71274) for 5 hours. Luciferase activity was measured using the ONE-Step™ Luciferase Assay System (BPS Bioscience #60690). Cells transduced with V γ 4V δ 1 TCR Lentivirus can be activated by the V δ 1 antibody, but not the V δ 2 antibody, while cells transduced with V γ 9V δ 2 TCR Lentivirus can be activated by the V δ 2 antibody, but not the V δ 1 antibody. CD3 agonist antibody treatment was run in parallel as a positive control.

Data shown is representative. For lot-specific information, please contact BPS Bioscience, Inc. at support@bpsbioscience.com.

Notes

To generate a V γ 9V δ 2 TCR expressing stable cell line, remove the growth medium 48 hours after transduction and replace it with fresh growth medium containing the appropriate amount of puromycin (as pre-determined from a killing curve, <https://bpsbioscience.com/cell-line-faq>), for antibiotic selection of transduced cells, followed by clonal selection.

Troubleshooting Guide

Visit bpsbioscience.com/lentivirus-faq for detailed troubleshooting instructions. For further questions, please email support@bpsbioscience.com.

References

Allison T. and Garboczi D., 2002 *Molecular Immunology* 38 (14): 1051-1061.
Sawaisorn P., et al., 2024 *Scientific Reports* 14: 1291.

Related Products

Products	Catalog #	Size
V γ 4V δ 1 TCR Lentivirus	78986	100 μ l/2 x 500 μ l

Version 042524