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Data Sheet
EZH2 (A677G) Chemiluminescent Assay Kit
Catalog # 52070
Size: 384 reactions

DESCRIPTION: The *EZH2 (A677G) Chemiluminescent Assay Kit* is designed to measure activity of the mutant EZH2 complex (EZH2 (A677G)/EED/SUZ12/RbAp48/AEBP) for screening and profiling purposes. The *EZH2 (A677G) Chemiluminescent Assay Kit* comes in a convenient format, with wells precoated with histone H3 peptide substrate, an antibody against methylated K27 residue of Histone H3, the secondary HRP-labeled antibody, S-adenosylmethionine, methyltransferase assay buffer, and purified EZH2 (A677G) complex for 384 enzyme reactions. The key to the EZH2 (A677G) Assay Kit is a highly specific antibody that recognizes methylated Histone H3K27. With this kit, only three simple steps on a microtiter plate are required for methyltransferase detection. First, S-adenosylmethionine is incubated with a sample containing assay buffer and methyltransferase enzyme for one hour. Next, primary antibody is added. Finally, the strips are treated with an HRP-labeled secondary antibody followed by addition of the HRP substrate to produce chemiluminescence that can then be measured using a chemiluminescence reader.

COMPONENTS:

| Catalog # | Component | Amount | Storage | |
|-----------|--|----------|---------|---|
| 51037 | EZH2 (A677G)/EED/SUZ12/RbAp48/AEBP2 | 80 µg | -80°C | Avoid freeze/ thaw cycles! |
| 52120 | 60 µM S-adenosylmethionine | 4x250 µl | -80°C | |
| 52140X | Primary Antibody 24 | 25 µl | -80°C | |
| 52131H | Secondary HRP-labeled Antibody 2 | 20 µl | -80°C | |
| 52170B | 4x HMT Assay Buffer 2B | 2x3 ml | -20°C | |
| 52100 | Blocking Buffer | 2x50 ml | +4°C | |
| | HRP chemiluminescent substrate A (translucent bottle) | 2x6 ml | +4°C | |
| | HRP chemiluminescent substrate B (brown bottle) | 2x6 ml | +4°C | |
| | 384-well plate precoated with histone substrate | 1 plate | +4°C | |

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MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:

TBST buffer (1x Tris-buffered saline, pH 8.0, containing 0.05% Tween-20)
Luminometer or microplate reader capable of reading chemiluminescence
Rotating or rocker platform

APPLICATIONS: Great for studying enzyme kinetics and HTS applications.

STABILITY: One year from date of receipt when stored as directed.

REFERENCE(S):

1. Dillon, S.C., *et al. Genome Biology* 2005; **6**:227.
2. Morin, R.D., *et al. Nat Genet.* 2010, **42**(2):181.

ASSAY PROTOCOL:

All samples and controls should be tested in duplicate.

Step 1:

- 1) Rehydrate the microwells by adding 90 μ l of TBST buffer (1x TBS, pH 8.0, containing 0.05% Tween-20) to every well. Incubate 15 minutes at room temperature. Tap the plate onto clean paper towels to remove liquid.
- 2) Thaw **60 μ M S-adenosylmethionine** on ice. Upon first thaw, briefly spin tube containing **60 μ M S-adenosylmethionine** to recover full content of the tube. Aliquot **60 μ M S-adenosylmethionine** into single use aliquots. Store remaining **60 μ M S-adenosylmethionine** in aliquots at -80°C immediately. *Note: S-adenosylmethionine is very sensitive to freeze/thaw cycles. Avoid multiple freeze-thaw cycles.*
- 3) Prepare the master mixture: N wells \times (7.5 μ l **4x HMT Assay Buffer 2B** + 2.5 μ l **60 μ M S-adenosylmethionine** + 15 μ l **H₂O**)

| | Blank | Substrate Control | Positive Control | Test Inhibitor |
|---------------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|
| 4x HMT assay buffer 2B | 7.5 μ l | 7.5 μ l | 7.5 μ l | 7.5 μ l |
| 60 μ M S-adenosylmethionine | 2.5 μ l | - | 2.5 μ l | 2.5 μ l |
| H ₂ O | 15 μ l | 17.5 μ l | 15 μ l | 15 μ l |
| Test Inhibitor/Activator | - | - | - | 5 μ l |
| Inhibitor buffer (no inhibitor) | 5 μ l | 5 μ l | 5 μ l | - |
| 1x HMT assay buffer 2B | 20 μ l | - | - | - |
| EZH2(A677G) (4-8 ng/ μ l) | - | 20 μ l | 20 μ l | 20 μ l |
| Total | 50 μl | 50 μl | 50 μl | 50 μl |

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- 4) Add 25 μ l of master mixture to each well designated for the "Positive Control", "Test Inhibitor", and "Blank". For the "Substrate Control", add 7.5 μ l **4x HMT Assay Buffer 2** + 17.5 μ l **H₂O**
- 5) Add 5 μ l of inhibitor solution of each well designated "Test Inhibitor". For the "Positive Control", "Substrate Control" and "Blank", add 5 μ l of the same solution without inhibitor (inhibitor buffer).
- 6) Add 20 μ l of **1 x HMT assay buffer 2B** to the well designated "Blank".
- 7) Thaw **EZH2(A677G) enzyme** on ice. Upon first thaw, briefly spin tube containing enzyme to recover full content of the tube. Aliquot **EZH2(A677G) enzyme** into single use aliquots. Store remaining undiluted enzyme in aliquots at -70°C immediately. *Note: **EZH2(A677G) enzyme** is very sensitive to freeze/thaw cycles. Do not re-use thawed aliquots or diluted enzyme.*
- 8) Dilute **EZH2(A677G) enzyme** in **1x HMT assay buffer 2B** at 4-8 ng/ μ l (80-160 ng/20 μ l). Keep diluted enzyme on ice until use. Discard any unused diluted enzyme after use.
- 9) Initiate reaction by adding 20 μ l of diluted **EZH2(A677G)** prepared as described above. Incubate at room temperature for one hour.
- 10) Wash the strips three times with 90 μ l TBST buffer. Blot dry onto clean paper towels.
- 11) Add 50 μ l of **Blocking Buffer** to every well. Shake on a rotating platform for 10 min. Remove supernatant as described above.

Step 2:

- 1) Dilute **Primary antibody 24** 800-fold with **Blocking Buffer**.
- 2) Add 50 μ l per well. Incubate 1 hour at room temperature with slow shaking.
- 3) Wash the strips three times with TBST buffer and incubate in **Blocking Buffer** as described in steps 1-10 and 1-11.

Step 3:

- 1) Dilute **Secondary HRP-labeled antibody 2** 1,000-fold with **Blocking Buffer**.
 - 2) Add 50 μ l per well. Incubate for 30 min. at room temperature with slow shaking.
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- 3) Wash the strips with TBST buffer and incubate in **Blocking Buffer** as described in step 1-10 and 1-11.
- 4) Just before use, mix on ice 25 μ l **HRP chemiluminescent substrate A** and 25 μ l **HRP chemiluminescent substrate B** and add 50 μ l per well. Discard any unused chemiluminescent reagent after use.
- 5) Immediately read sample in a luminometer or microtiter-plate capable of reading chemiluminescence. "Blank" value is subtracted from all readings.

Reading Chemiluminescence:

Chemiluminescence is the emission of light (luminescence) which results from a chemical reaction. The detection of chemiluminescence requires no wavelength selection because the method used is emission photometry and is not emission spectrophotometry.

To properly read chemiluminescence, make sure the plate reader is set for LUMINESCENCE mode. Typical integration time is 1 second, delay after plate movement is 100 msec. Do not use a filter when measuring light emission. Typical settings for the Synergy 2 BioTek plate reader are: use the "hole" position on the filter wheel; Optics position: Top; Read type: endpoint. Sensitivity may be adjusted based on the luminescence of a control assay without enzyme (typically we set this value as 100).

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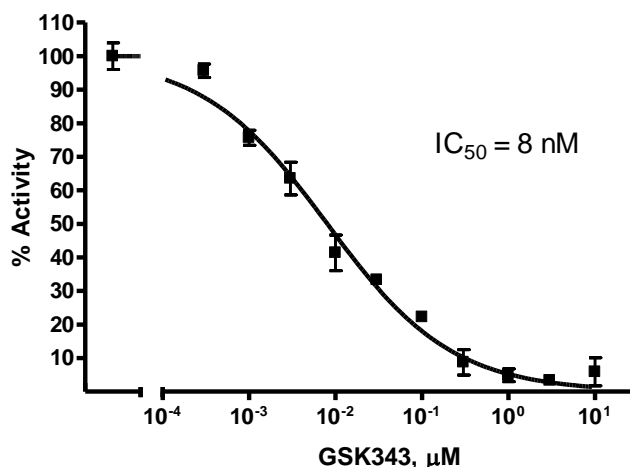
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Example of Assay Results:

EZH2 A677 Activity



EZH2(A677G) enzyme activity, measured using the *EZH2 (A677G) Chemiluminescent Assay Kit*, BPS Bioscience # 52070. Luminescence was measured using a Bio-Tek fluorescent microplate reader. *Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at info@bpsbioscience.com.*

RELATED PRODUCTS

| <u>Product Name</u> | <u>Catalog #</u> | <u>Size</u> |
|---|------------------|-------------|
| EZH2 (A677G)/EED/SUZ12/RbAp48/AEBP2 | 51037 | 20 µg |
| EZH2 (Y641F)/EED/SUZ12/RbAp48/AEBP2 | 51017 | 20 µg |
| EZH2 (Y641C)/EED/SUZ12/RbAp48/AEBP2 | 51029 | 20 µg |
| EZH2 (Y641N)/EED/SUZ12/RbAp48/AEBP2 | 51028 | 20 µg |
| EZH2 (Y641S)/EED/SUZ12/RbAp48/AEBP2 | 51013 | 20 µg |
| EZH2 (Y641H)/EED/SUZ12/RbAp48/AEBP2 | 51011 | 20 µg |
| EZH2/EED/SUZ12/RbAp48/AEBP2 | 51004 | 50 µg |
| EZH2 Chemiluminescent Assay Kit | 52009L | 96 rxns. |
| EZH2 (Y641F) Chemiluminescent Assay Kit | 52075 | 96 rxns. |
| EZH2 (Y641N) Chemiluminescent Assay Kit | 52076 | 96 rxns. |
| EZH2 (A738T) Chemiluminescent Assay Kit | 52067 | 96 rxns. |
| EZH2 Homogeneous Assay Kit | 52059 | 384 rxns. |
| EZH2 (Y641N) TR-FRET Assay Kit | 52078 | 384 rxns. |

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TROUBLESHOOTING GUIDE

| Problem | Possible Cause | Solution |
|--|---|--|
| Luminescence signal of positive control reaction is weak | EZH2 (A677G) Complex has lost activity | Enzyme loses activity upon repeated freeze/thaw cycles. Use fresh EZH2 (A677G) Complex, BPS Bioscience #51037. Store enzyme in single-use aliquots. Increase time of enzyme incubation. Increase enzyme concentration. |
| | Antibody reaction is insufficient | Increase time for primary antibody incubation. Avoid freeze/thaw cycles of antibodies. |
| | Incorrect settings on instruments | Refer to instrument instructions for settings to increase sensitivity of light detection. See section on "Reading Chemiluminescence" above. |
| | Chemiluminescent reagents mixed too soon | Chemiluminescent solution should be used within 15 minutes of mixing. Ensure both reagents are properly mixed. |
| Luminescent signal is erratic or varies widely among wells | Inaccurate pipetting/technique | Run duplicates of all reactions. Use a multichannel pipettor. Use master mixes to minimize errors. |
| | Bubbles in wells | Pipette slowly to avoid bubble formation. Tap plate lightly to disperse bubbles; be careful not to splash between wells. |
| Background (signal to noise ratio) is high | Insufficient washes | Be sure to include blocking steps after wash steps. Increase number of washes. Increase wash volume. Increase Tween-20 concentration to 0.1% in TBST. |
| | Sample solvent is inhibiting the enzyme | Run negative control assay including solvent. Maintain DMSO level at <1%. Increase time of enzyme incubation. |
| | Results are outside the linear range of the assay | Use different concentrations of EZH2 (A677G) Complex, BPS #51037 to create a standard curve. |

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