

**Data Sheet**  
**JARID1B Homogeneous Assay Kit**  
**Catalog #50512-2**  
**Size : 384 reactions**

**DESCRIPTION:** The *JARID1B Homogeneous Assay Kit* is designed to measure JARID1B activity for screening and profiling applications. JARID1B, also known as PLU-1 and KDM5B, is a JumonjiC (JmjC) and ARID domain-containing histone lysine demethylase that exhibits demethylation activity toward H3-K<sub>4</sub>Me<sup>2</sup> and H3-K<sub>4</sub>Me<sup>3</sup>. The *JARID1B Homogeneous Assay Kit* comes in a convenient AlphaLISA<sup>®</sup> format (Scheme 1), with biotinylated histone H3 peptide substrate, primary antibody, demethylase assay buffer, and purified JARID1B for 384 enzyme reactions. The key to the *JARID1B Homogeneous Assay Kit* is a highly specific antibody that recognizes demethylated substrate. With this kit, only three simple steps on a microtiter plate are required for methyltransferase detection. First, a sample containing JARID1B enzyme is incubated with the biotinylated substrate. Next, acceptor beads and primary antibody are added, then donor beads, followed by reading the Alpha-counts.

**COMPONENTS:**

Catalog #	Component	Amount	Storage	
50121	JARID1B (KDM5B)	60 µg	-80°C	<b>(Avoid freeze/thaw cycles!)</b>
52140M4	Primary antibody 13-4	20 µl	-80°C	
79840	Biotinylated histone H3 peptide substrate	500 rxns	-80°C	
52407	4x HDM Assay Buffer 2	3 ml	-80°C	
	4x Detection buffer D	2 ml	-20°C	

**MATERIALS REQUIRED BUT NOT SUPPLIED:**

AlphaLISA<sup>®</sup> anti-rIgG acceptor beads, 5 mg/ml (PerkinElmer #AL104C)  
 AlphaScreen<sup>®</sup> Streptavidin-conjugated donor beads, 5 mg/ml (PerkinElmer #6760002S)  
 Optiplat -384 (PerkinElmer #6007290)  
 AlphaScreen<sup>®</sup> microplate reader  
 Adjustable micropipettor and sterile tips

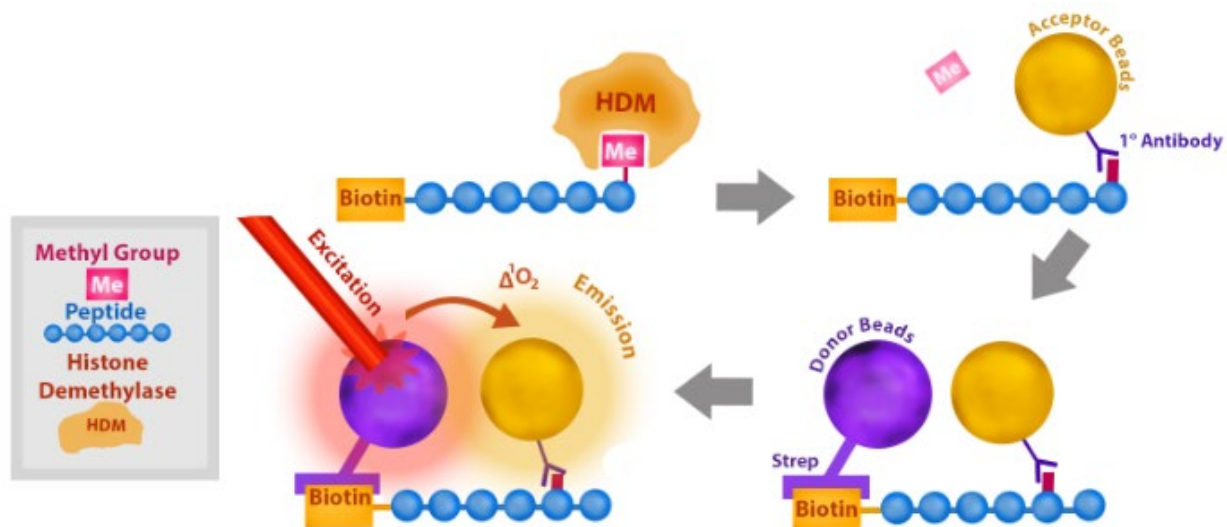
**APPLICATIONS:** Great for studying enzyme kinetics and HTS applications.

**SAFETY:** This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous. Do not ingest, inhale, get in eyes, on skin, or on clothing. If so, wash thoroughly.

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**Scheme 1:** Our histone demethylase assays utilize highly specific antibodies that recognize demethylated products. First, a sample containing the enzyme is incubated with a biotinylated substrate. Next, acceptor beads and primary antibody are added, then donor beads, followed by reading the Alpha-counts, as shown below.



**CONTRAINDICATIONS:** Green and blue dyes that absorb light in the AlphaScreen signal emission range (520-620 nm), such as Trypan Blue. Avoid the use of the potent singlet oxygen quenchers such as sodium azide ( $\text{NaN}_3$ ) or metal ions ( $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}$  and  $\text{Ni}^{2+}$ ). The presence of >1% RPMI 1640 culture medium leads to a signal reduction due to the presence of excess biotin and iron in this medium. MEM, which lacks these components, does not affect AlphaScreen assays.

**STABILITY:** At least one year from date of receipt when stored as directed.

**REFERENCE(S):**

1. Lahoud, M.H., *et al. Genome Res* **11** (8): 1327–34.

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6042 Cornerstone Court W, Ste B  
San Diego, CA 92121  
**Tel:** 1.858.202.1401  
**Fax:** 1.858.481.8694  
**Email:** [info@bpsbioscience.com](mailto:info@bpsbioscience.com)

### ASSAY PROTOCOL:

All samples and controls should be tested in duplicate. We recommend preincubating the enzyme with inhibitor, however, it is acceptable to add the substrate mixture and inhibitor followed by diluted JARID1B without the preincubation step.

#### Step 1:

- 1) Re-suspend lyophilized **Biotinylated histone H3 peptide substrate** in 500  $\mu$ l of distilled water.
- 2) Prepare serial dilutions of the test inhibitors in **1x HDM Assay Buffer 2** (Scheme 2). Add 3  $\mu$ l of inhibitor solution to each well designated "Test Sample". For the wells designated "Blank" and "Positive Control" add 3  $\mu$ l of the same solution without inhibitor (typically **1x HDM Assay Buffer 2** with respective concentration of DMSO).
- 3) Thaw **JARID1B** on ice. Upon first thaw, briefly spin tube containing enzyme to recover full content of the tube. Aliquot **JARID1B** enzyme into single use aliquots. Store remaining undiluted enzyme in aliquots at  $-80^{\circ}\text{C}$  immediately. *Note: **JARID1B** is very sensitive to freeze/thaw cycles. Do not re-use thawed aliquots or diluted enzyme.*
- 4) Dilute **JARID1B** in **1x HDM Assay Buffer 2** at 25 ng/ $\mu$ l (100 ng/4  $\mu$ l). Keep diluted enzyme on ice until use. Discard any unused diluted enzyme after use.
- 5) Preincubate 4  $\mu$ l of diluted **JARID1B** with 3  $\mu$ l of diluted inhibitor(s) for up to 30 minutes at room temperature, with slow shaking. For the wells designated as "Blank", add 4  $\mu$ l **1x HDM Assay Buffer 2**.
- 6) Prepare master mix: N wells  $\times$  (1.5  $\mu$ l **4x HDM Assay Buffer 2** + 1  $\mu$ l **Biotinylated substrate** + 0.5  $\mu$ l **distilled water**).
- 7) Initiate reaction by adding 3  $\mu$ l of master mix prepared as described above. Incubate at room temperature for one hour. *Note: All incubations are done with slow shaking on a rotator platform.*

**Scheme 2:** The serial dilution of the compounds was first performed in 100% DMSO with the highest concentration at (X) mM. Each intermediate compound dilution (in 100% DMSO) will then get directly diluted 30x fold into **1x HDM Assay Buffer 2** for 3.3x concentration (DMSO). From this intermediate step, 3  $\mu$ l of compound is added to 4  $\mu$ l of demethylase enzyme dilution is incubated for 30 minutes at room temperature. After this incubation, 3  $\mu$ l of peptide substrate is added. The final DMSO concentration is 1% for all wells.

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Reagent	Blank	Positive Control	Test Inhibitor
1x HDM Assay Buffer 2	4 $\mu$ l	–	–
4x HDM Assay Buffer 2	1.5 $\mu$ l	1.5 $\mu$ l	1.5 $\mu$ l
Biotinylated Substrate	1 $\mu$ l	1 $\mu$ l	1 $\mu$ l
Distilled water	0.5 $\mu$ l	0.5 $\mu$ l	0.5 $\mu$ l
Test Inhibitor/Activator	–	–	3 $\mu$ l
1x HDM Assay Buffer 2 (3.3% DMSO)	3 $\mu$ l	3 $\mu$ l	–
JARID1B (25 ng/ $\mu$ l)	–	4 $\mu$ l	4 $\mu$ l
<b>Total</b>	<b>10 <math>\mu</math>l</b>	<b>10 <math>\mu</math>l</b>	<b>10 <math>\mu</math>l</b>

**Step 2:**

**Note: Protect your samples from direct exposure to light!**

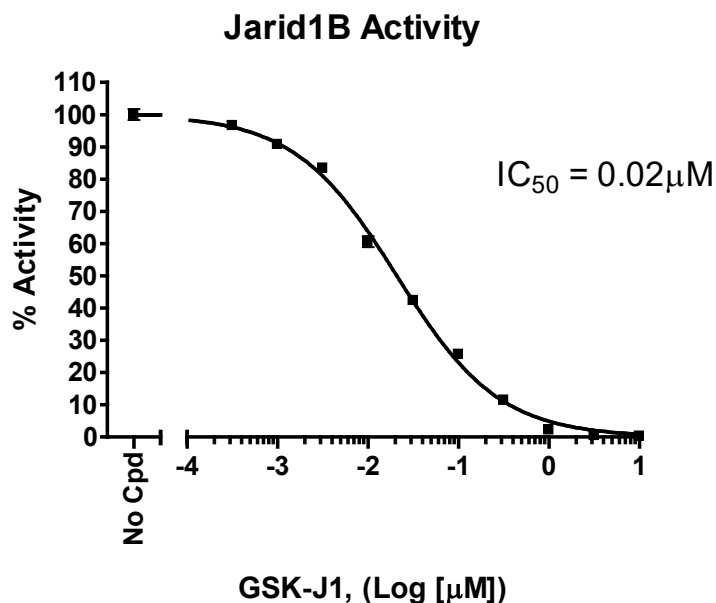
- 1) Dilute anti-Rabbit Acceptor beads (PerkinElmer #AL104C) (1:500) and Primary antibody 13-4 (1:200) with 1x Detection buffer in one step. Add 10  $\mu$ l of acceptor beads/antibody mixture per well. Incubate 30 min at room temperature.

**Step 3:**

- 1) Dilute **Streptavidin-conjugated donor beads** (PE #6760002S) 125-fold with **1x Detection buffer**. Add 10  $\mu$ l of donor beads per well. Shake on a rotator platform for 30 minutes at room temperature.
- 2) Read Alpha-counts.

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**Example of Assay Results:**


JARID1B enzyme activity, measured using the *JARID1B Homogeneous Assay Kit*, BPS Bioscience Cat. #50512. Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at [info@bpsbioscience.com](mailto:info@bpsbioscience.com)

**RELATED PRODUCTS:**

<u>Product Name</u>	<u>Catalog #</u>	<u>Size</u>
JARID1A Assay Kit, Homogeneous	50510	384 reactions
JARID1C Assay Kit, Homogeneous	50511	384 reactions
JARID1B Assay Kit, TR-FRET	50514	384 reactions
JARID1A Assay Kit, Chemiluminescence	50513	96 reactions
JARID1B Assay Kit, Chemiluminescence	50517	96 reactions
JARID1A recombinant protein	50110	20 $\mu$ g
JARID1B recombinant protein	50121	20 $\mu$ g
Mouse JARID1B recombinant protein	50122	20 $\mu$ g
JARID1C recombinant protein	50112	20 $\mu$ g

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### TROUBLESHOOTING GUIDE

Problem	Possible Cause	Solution
Alpha-counts signal of positive control reaction is same as "blank" value.	JARID1B has lost activity	Enzyme loses activity upon repeated freeze/thaw cycles. Use fresh JARID1A, BPS Bioscience #50121. Store enzyme in single-use aliquots. Increase time of enzyme incubation. Increase enzyme concentration.
	Streptavidin Donor beads or anti-rlgG acceptor beads fail to show significant signal.	Reorder Streptavidin Donor beads or anti-rlgG acceptor beads from Perkin Elmer.
	Incorrect settings on instruments	Refer to instrument instructions for correct settings to increase sensitivity of light detection.
Alpha-counts signal is erratic or varies widely among wells	Inaccurate pipetting/technique	Run duplicates of all reactions. Use a multichannel pipettor. Use master mixes to minimize errors.

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