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Data Sheet

IFN- γ (Human) Colorimetric ELISA Detection Kit

Catalog #79777-1

Size: 96 reactions

DESCRIPTION: IFN- γ is a cytokine that is primarily secreted by activated T cells and natural killer (NK) cells and plays an important role in activating innate/adaptive immunity. It also has been recognized as an indicator of T cell activation suggesting IFN- γ activation may be a prerequisite for CAR-T cell activity. The *IFN- γ (Human) Colorimetric ELISA Detection Kit* is designed for detecting and quantifying human interferon- γ in cell culture medium. This kit comes in a convenient 96-well format, with capture and detection antibodies for IFN- γ , streptavidin-labeled HRP, blocking buffer, IFN- γ standard, and colorimetric HRP substrate for a 96-well plate. Only a few simple steps on a microtiter plate are required for the assay. First, the capture antibody is coated on a 96-well plate. Next, samples containing IFN- γ are incubated on the coated plate followed by detecting the captured IFN- γ with the detection antibody. Finally, the plate is treated with streptavidin-HRP followed by addition of a colorimetric HRP substrate to produce color, which can then be measured using a UV/Vis spectrophotometer microplate reader.

COMPONENTS:

Catalog #	Component	Amount	Storage	
	Interferon- γ capture antibody	10 μ l	-80°C	(Avoid freeze/thaw cycles!)
	Interferon- γ detection antibody, biotinylated	5 μ l	-80°C	
	Human IFN- γ standard (1 μ g/ml)	20 μ l	-80°C	
80611	Streptavidin-HRP	5 μ l	+4°C	
79743	Blocking Buffer 3	50 ml	+4°C	
79651	Colorimetric HRP substrate	10 ml	+4°C	
79964	Transparent 96-well microplate	1	+4°C	

MATERIALS OR INSTRUMENTS REQUIRED BUT NOT SUPPLIED:

PBS (Phosphate-buffered saline)

PBST (PBS with 0.05% Tween-20)

1N HCl (aqueous)

Rotating or rocker platform

UV/Vis spectrophotometer microplate reader capable of reading absorbance at 450 nm*

*Alternatively, a spectrophotometer reading at 650 nm may be used without adding 1N HCl, but sensitivity of the assay will be reduced.

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APPLICATIONS: This kit is useful for cytokine detection in cell culture medium.

STABILITY: One year from date of receipt when stored as directed.

REFERENCES:

1. Billiau, A., 1996. *Interferon gamma: biology and role in pathogenesis*, Adv. Immunology, **62:** 61-130

ASSAY PROTOCOL:

All samples and standards should be tested in duplicate.

Coating the plate with Capture Antibody:

- 1) Thaw **Capture Antibody** on ice. Upon first thaw, *briefly* spin tube containing **Capture Antibody** to recover the full contents of the tube.
- 2) Dilute **Capture Antibody** to 2 ng/ μ l in ~ 5 mL PBS for a 96-well plate.
- 3) Add 50 μ l of diluted **Capture Antibody** solution to each well and incubate overnight at 4°C. (Remaining **Capture Antibody** can be stored at 4°C)

**After overnight coating, it is highly recommended that all remaining steps are completed the following day to obtain optimal sensitivity.*

- 4) After the overnight incubation, decant to remove the solution. Wash the plate 2 times with 200 μ l/well of PBS with 0.05% Tween-20 (PBST). Tap plate onto clean paper towels to remove liquid.
- 5) Block wells by adding 200 μ l of **Blocking Buffer** to each well. Incubate for 1 hour at room temperature. Decant to remove the blocking buffer and wash the plate 2 times with 200 μ l/well of PBST. Tap plate onto clean paper towels to remove liquid.

Step 1:

- 1) Prepare the sample by diluting with the **Blocking Buffer**. The detection range of the *Colorimetric Human IFN- γ Detection Kit* is 5 pg/ml – 500 pg/ml (**Figure 1**). Roughly estimate the amount of human IFN- γ in the sample and dilute it accordingly. For quantification, **human IFN- γ standard** can be serially diluted (500 pg/ml to 5 pg/ml) in **Blocking Buffer** and run in the same plate. (Aliquot remaining **human IFN- γ standard** into single use aliquots. Immediately store remaining undiluted protein in aliquots at -80°C.)

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- 2) Add 50 μ l of the diluted samples (or **human IFN- γ standard**) to each well and incubate the plate for 2 hours at room temperature.
- 3) After 2 hours incubation, decant to remove the solution and wash the plate 2 times with 200 μ l/well of PBST. Tap plate onto clean paper towels to remove liquid.
- 4) Dilute **biotinylated-detection Antibody** 1:1,000 in the Blocking Buffer, and add 50 μ l to each well. Incubate the plate for 1 hour at room temperature.
- 5) After 1-hour incubation, decant to remove the solution and wash the plate 3 times with 200 μ l/well of PBST. Tap plate onto clean paper towels to remove liquid.
- 6) Dilute **Streptavidin-HRP** 1:1,000 in the Blocking Buffer, and add 50 μ l to each well. Incubate the plate for 30 minutes at room temperature.
- 7) After 30 minutes incubation, decant to remove the solution and wash the plate 5 times with 200 μ l/well of PBST. Tap plate onto clean paper towels to remove liquid
- 8) Add 100 μ l of the **Colorimetric HRP substrate** to each well and incubate the plate at room temperature until blue color is developed in the positive control well. This usually takes several minutes to fully develop. The optimal incubation time may vary, and should be determined empirically by the user.
- 9) After the blue color is developed, add 100 μ l of 1 M HCl to each well. Read the absorbance at 450 nm using UV/Vis spectrophotometer microplate reader. The blank wells should exhibit an absorbance of \sim 0.05 at 450 nm. *Alternatively, the plate may be read at 650 nm without adding 1N HCl, but the Signal-to-Background ratio will be decreased.*

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Example of Detection Results:

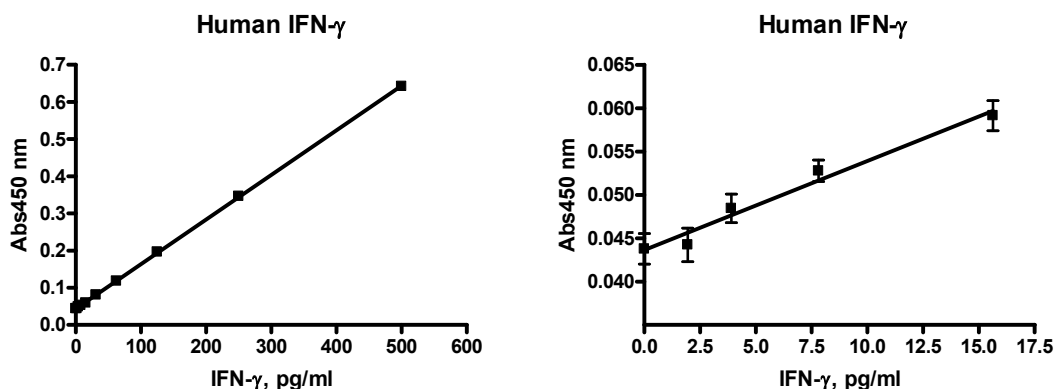


Figure 1. Detection of the human IFN- γ (left: 2 pg/ml – 500 pg/ml, right: 2 pg/ml – 15 pg/ml) standard using the *Colorimetric Human IFN- γ Detection Kit*. Data shown is lot-specific. For lot-specific information, please contact BPS Bioscience, Inc. at info@bpsbioscience.com

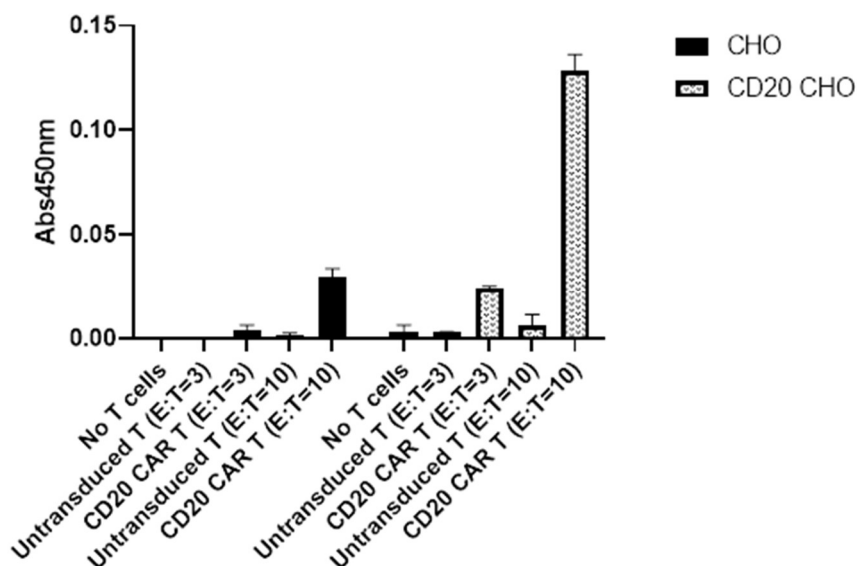


Figure 2. IFN- γ production of anti-CD20 CAR-T cells. T-cells were isolated from PBMC (BPS Bioscience #79059) and transduced with the anti-CD20 CAR-T lentivirus (BPS Bioscience #78606) and incubated with the target cells (CD 20 expressing Luciferase CHO cells, BPS Bioscience #79624-H) for 24 hours. Cell growing medium was 3-folds diluted in the Blocking buffer and amount of the secreted IFN- γ was measured by the Colorimetric Human IFN- γ Detection Kit. Absorbance from the medium-only well was subtracted.

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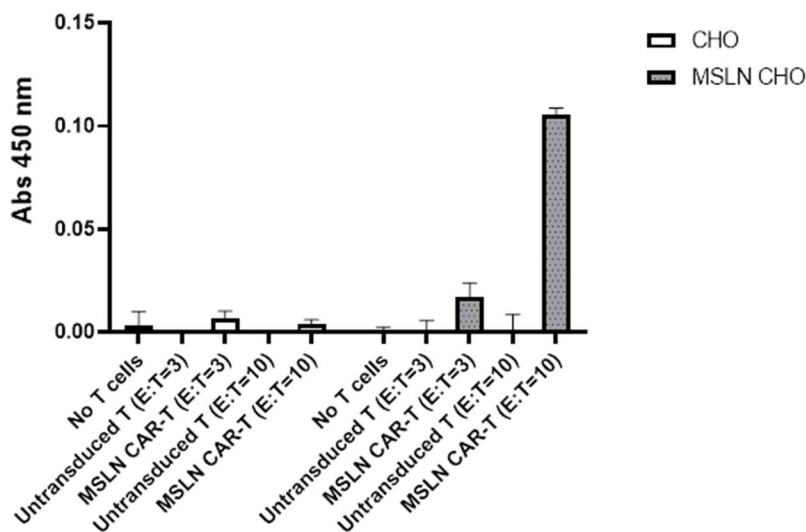


Figure 3. IFN- γ production of anti-Mesothelin CAR-T cells. T-cells were isolated from PBMC (BPS Bioscience #79059) and transduced with the anti-Mesothelin CAR-T lentivirus (BPS Bioscience #78703) and incubated with the target cells (Mesothelin expressing CHO cells, BPS Bioscience #78132) for 24 hours. Cell growing medium was 3-folds diluted in the Blocking buffer and amount of the secreted IFN- γ was measured by the Colorimetric Human IFN- γ Detection Kit. Absorbance from the medium-only well was subtracted.

RELATED PRODUCTS:

<u>Product Name</u>	<u>Catalog #</u>	<u>Size</u>
Human Interleukin-1 gamma	90162-A	20 μ g
Human Interleukin-1 gamma	90162-B	100 μ g
Mouse Interleukin-1 gamma	90163-A	20 μ g
Mouse Interleukin-1 gamma	90163-B	100 μ g
Rat Interleukin-1 gamma	90164-A	20 μ g
Rat Interleukin-1 gamma	90164-B	100 μ g
Streptavidin-HRP	80611	100 μ l
Blocking Buffer 3	79743	50 ml
IFN- α (Human) Colorimetric ELISA Detection Kit	79779	96 rxns.
IFN- β (Human) Colorimetric ELISA Detection Kit	79780	96 rxns.

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TROUBLESHOOTING GUIDE

Problem	Possible Cause	Solution
Colorimetric signal of positive control reaction is weak	Antibodies have lost activity	Antibodies and IL-1 γ standard may lose activity upon repeated freeze/thaw cycles. Use fresh protein. Store proteins in single-use aliquots. Increase time of incubation. Increase protein or antibody concentration.
	Incorrect settings on instruments	Refer to instrument instructions for settings to increase sensitivity.
	Colorimetric HRP substrate was not incubated long enough	Increase the amount of time that the colorimetric HRP substrate is incubated in the wells. Avoid azides.
Colorimetric signal is erratic or varies widely among wells	Inaccurate pipetting/technique	Run duplicates of all reactions. Use a multichannel pipettor. Use master mixes to minimize errors.
	Bubbles in wells	Pipette slowly to avoid bubble formation. Tap plate lightly to disperse bubbles; be careful not to splash between wells.
	Signal is out of range of detection (too high)	Decrease the amount of time that the colorimetric HRP substrate is incubated in the wells
Background (signal to noise ratio) is high	Insufficient washes or blocking	Be sure to include blocking steps after wash steps. Increase number of washes. Increase wash volume. Increase Tween-20 concentration to 0.1% in TBST. Be sure to dilute Streptavidin-HRP in blocking buffer, not assay buffer.
	Sample solvent is inhibiting the protein	Run negative control assay including solvent. Maintain DMSO level at <1%. Increase time of protein incubation.
	Results are outside the linear range of the assay	Use different concentrations of IL-1 γ standard to create a standard curve.

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