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Data Sheet

Empty vector control - HEK293 Recombinant Cell line
Catalog #: 90334

Product description

Recombinant HEK293 cell transfected with empty expression vector containing the neomycin resistance gene. This cell line serves as a negative control for

- TMEM16A-HEK293 Cell Line Cat. #90230
- TRPC3-HEK293 Cell Line Cat. #90130
- TRPC7-HEK293 Cell Line Cat. #90030
- IKCA1 (KCNN4) - HEK293 Cell Line Cat. #90330
- ERG (Kv11.1) - HEK293 Cell Line Cat. #60619

Format
Each vial contains 1.5 X 10^6 cells in 1 ml of 10% DMSO.

Storage
Immediately upon receipt, store in liquid nitrogen.

Mycoplasma Testing
The cell line has been screened using the PCR-based Venor™GeM Mycoplasma Detection kit (Sigma-Aldrich) to confirm the absence of Mycoplasma species.

Culture conditions

Thaw Medium 1 (BPS Cat. #60187): MEM medium (Hyclone #SH30024.01) supplemented with 10% FBS (Invitrogen #26140-079), 1% non-essential amino acids (Hyclone #SH30238.01), 1 mM Na pyruvate (Hyclone #SH30239.01), 1% Penicillin/Streptomycin (Hyclone SV30010.01)

Growth Medium 1B (BPS Cat. #79531): Thaw Medium 1 (BPS Cat. #60187) plus 400 μg/ml of Geneticin (life Technologies #11811031) to ensure the recombinant expression is maintained.

Cells should be grown using at 37°C with 5% CO_2 using Growth Medium 1B. hERG-HEK293 cells should exhibit a typical cell division time of ~24 hours.

To thaw the cells, it is recommended to quickly thaw the frozen cells from liquid nitrogen in a 37°C water-bath and transfer to a tube containing 10 ml of Thaw Medium 1 **(no Geneticin)**. Spin down cells, resuspend cells in pre-warmed Thaw Medium 1 **(no Geneticin)**, and transfer resuspended cells to T25 flask. Culture at 37°C in a CO_2 incubator overnight. The next day, replace the medium with fresh Thaw Medium 1 **(no Geneticin)** and continue growing culture in

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a CO_2 incubator at 37°C until the cells are ready to be split. Cells should be split before they reach complete confluence. At first passage, switch to Growth Medium 1B **(contains Geneticin)**.

To passage the cells, rinse cells with phosphate buffered saline (PBS) and detach cells from culture vessel with 0.05% Trypsin/EDTA. Add Growth Medium 1B and transfer to a tube, spin down cells, resuspend cells, and seed appropriate aliquots of cell suspension into new culture vessels. Subcultivation ration: 1:10 to 1:20 weekly or twice a week.

To freeze cells, rinse cells with phosphate buffered saline (PBS) and detach cells from culture vessel with 0.05% Trypsin/EDTA. Add Growth Medium 1B and transfer to a tube, spin down cells and resuspend in freezing medium (90% FBS, 10% DMSO). Freeze cells at -80°C overnight and transfer to liquid nitrogen the next day.

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