

Description

The CRE/CREB Luciferase Reporter CHO Cell Line (cAMP/PKA Signaling Pathway) contains a firefly luciferase gene under the control of multimerized cAMP response element (CRE) stably integrated into CHO cells. Elevation of the intracellular cAMP level activates cAMP response element binding protein (CREB) to bind CRE and induces the expression of luciferase. This cell line is validated for response to Forskolin.

Background

Cyclic adenosine monophosphate (cAMP) is a second messenger involved in cell signaling that regulates various physiological and pathological processes. cAMP regulates the transcription of target genes by activating protein kinase A (PKA) and the transcription factor cAMP response element-binding protein (CREB), a downstream effector. CRE is the target of many extracellular and intracellular signaling pathways, including cAMP, calcium, GPCR (G-protein coupled receptors), and neurotrophins. In the cAMP/PKA signaling pathway, CREB is activated via PKA-mediated phosphorylation and binds to CRE with a general motif of 5'-TGACGTCA-3'. The cAMP/PKA/CREB signaling pathway has both tumor-suppressive and tumor-promoting effects in cancer cells and can be useful in studying cancer signaling pathways.

Application(s)

- Monitor cAMP/PKA signaling pathway activity.
- Screen for activators or inhibitors of cAMP/PKA signaling pathway.

Materials Provided

Components	Format
2 vials of frozen cells	Each vial contains 2×10^6 cells in 1 ml of cell freezing medium (BPS Bioscience #79796)

Parental Cell Line

CHO-K1 cells, Chinese Hamster Ovary, epithelial-like cells, adherent

Mycoplasma Testing

The cell line has been screened to confirm the absence of Mycoplasma species.

Materials Required but Not Supplied



These materials are not supplied with the cell line but are necessary for cell culture and cellular assays. BPS Bioscience's reagents are validated and optimized for use with this cell line and are highly recommended for best results. Media components are provided in the Media Formulations section below.

Media Required for Cell Culture

Name	Ordering Information
Thaw Medium 3	BPS Bioscience #60186
Growth Medium 3D	BPS Bioscience #79539

Storage Conditions



Cells are shipped in dry ice and should immediately be thawed or stored in liquid nitrogen upon receipt. Do not use a -80°C freezer for long term storage. Contact technical support at support@bpsbioscience.com if the cells are not frozen in dry ice upon arrival.

Media Formulations

For best results, it is *highly recommended* to use these validated and optimized media from BPS Bioscience. Other preparations or formulations of media may result in suboptimal performance.



Note: Thaw Media do *not* contain selective antibiotics. However, Growth Media *do* contain selective antibiotics, which are used for maintaining the presence of the transfected gene(s) over passages.

Cells should be grown at 37 °C with 5% CO₂. BPS Bioscience's cell lines are stable for at least 15 passages when grown under proper conditions.

Media Required for Cell Culture

Thaw Medium 3 (BPS Bioscience #60186):

F-12K medium supplemented with 10% FBS, 1% Penicillin/Streptomycin.

Growth Medium 3D (BPS Bioscience #79539):

F-12K medium supplemented with 10% FBS, 1% Penicillin/Streptomycin plus 1000 ug/ml of Geneticin.

Materials Required for Cellular Assay

Name	Ordering Information
Assay Medium: Thaw Medium 3	BPS Bioscience #60186
Growth Medium 3D	BPS Bioscience #79539
Forskolin	Sigma #F3917
96-well tissue culture treated white clear-bottom assay plate	Corning #3610
ONE-Step™ Luciferase Assay System	BPS Bioscience #60690
Luminometer	

Cell Culture Protocol**Cell Thawing**

- Swirl the vial of frozen cells for approximately 60 seconds in a 37°C water bath. As soon as the cells are thawed (it may be slightly faster or slower than 60 seconds), quickly transfer the entire contents of the vial to a tube containing 10 ml of pre-warmed Thaw Medium 3 (**no geneticin**).
Leaving the cells in the water bath at 37°C for too long will result in rapid loss of viability.
- Immediately spin down the cells at 300 x g for 5 minutes, remove the medium and resuspend the cells in 5 ml of pre-warmed Thaw Medium 3 (**no geneticin**).
- Transfer the resuspended cells to a T25 flask or T75 flask and incubate at 37°C in a 5% CO₂ incubator.
- After 24 hours of culture, check for cell attachment and viability. Change medium to fresh Thaw Medium 3 (**no geneticin**) and continue growing in a 5% CO₂ incubator at 37°C until the cells are ready to passage.
- Cells should be passaged before they are fully confluent. At first passage and subsequent passages, use Growth Medium 3D containing Geneticin (**G418**).

Cell Passage

- Aspirate the medium, wash the cells with phosphate buffered saline (PBS), and detach the cells from the culture vessel with 0.25% Trypsin/EDTA.

2. Once the cells have detached, add Growth Medium 3D and transfer to a tube. Spin down cells at 300 x g for 5 minutes, remove the medium and resuspend the cells in Growth Medium **3D (contains Geneticin)**. Seed into new culture vessels at the desired sub-cultivation ratio of 1:10 to 1:20 weekly or twice per week.

Cell Freezing

1. Aspirate the medium, wash the cells with phosphate buffered saline (PBS), and detach the cells from the culture vessel with 0.25% Trypsin/EDTA.
2. Once the cells have detached, add Growth Medium **3D** and count the cells.
3. Spin down the cells at 300 x g for 5 minutes, remove the medium and resuspend the cells in 4°C Freezing Medium (BPS Bioscience #79796, or 10% DMSO + 90% FBS) at $\sim 2 \times 10^6$ cells/ml.
4. Dispense 1 ml of cell aliquots into cryogenic vials. Place the vials in an insulated container for slow cooling and store at -80°C overnight.
5. Transfer the vials to liquid nitrogen the next day for storage.



Note: It is recommended to expand the cells and freeze at least 10 vials at an early passage for future use.

Validation Data

- The following assays are designed for 96-well format. To perform the assay in different tissue culture formats, cell number and reagent volumes should be scaled appropriately.
- The experiment should be performed in triplicates.

Dose response of Forskolin in CRE/CREB Luciferase Reporter CHO cells

1. Seed CRE/CREB Luciferase Reporter CHO cells at a density of $\sim 32,000$ cells per well into white clear-bottom 96-well microplate in 90 μ l of Thaw Medium 3.
2. Incubate cells at 37°C with 5% CO₂ incubator overnight.
3. Prepare a serial dilution of Forskolin in Thaw Medium 3, at concentrations 10-fold higher than the desired final concentrations. Add 10 μ l of diluted Forskolin to the “stimulated” wells.
4. Add 10 μ l of Thaw Medium 3 to the “unstimulated control” wells (for measuring uninduced level of CRE/CREB reporter activity).
5. Add 100 μ l of Thaw Medium 3 to cell-free control wells (for determining background luminescence).
6. Incubate at 37°C with 5% CO₂ for 5-6 hours.
7. Prepare ONE-Step™ Luciferase Assay reagent according to recommended instructions. Add 100 μ l of ONE-Step™ Luciferase Assay reagent per well. Incubate at room temperature for 15 to 30 minutes and measure luminescence using a luminometer.

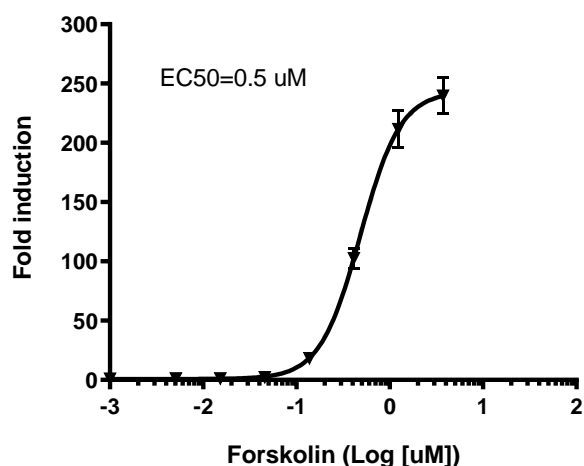


Figure 1. Forskolin dose response in CRE/CREB Luciferase Reporter CHO cells.

Cells were treated with increasing concentrations of Forskolin for ~ 5 hours. The results are shown as fold induction of luciferase reporter expression. Fold induction was determined by comparing values against the mean value for unstimulated control cells.

References

Kandel ER (2012). The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CPEB. *Mol Brain* 5: 14.

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Troubleshooting Guide

Visit bpsbioscience.com/cell-line-faq for detailed troubleshooting instructions. For all further questions, please email support@bpsbioscience.com.

Related Products

Products	Catalog #	Size
CRE/CREB reporter (Luc) - HEK293 Cell line	60515	2 vials
PDE7A/CRE Reporter - HEK293 Recombinant Cell Line	60413	2 vials
CRE/CREB Reporter Kit (cAMP/PKA Signaling Pathway)	60611	500 reactions
Transfection Collection™: CRE/CREB Transient Pack (cAMP/PKA Cell Signaling Pathway)	79267	100 reactions