

## Description

The HDAC2 Fluorogenic Assay Kit is designed to measure HDAC2 (histone deacetylase 2) activity for screening and profiling applications. The assay kit comes in a convenient 96-well format, with enough purified recombinant HDAC2 enzyme, fluorogenic substrate, HDAC Developer and assay buffer for 100 enzyme reactions. This kit also contains the inhibitor Trichostatin A as a control.

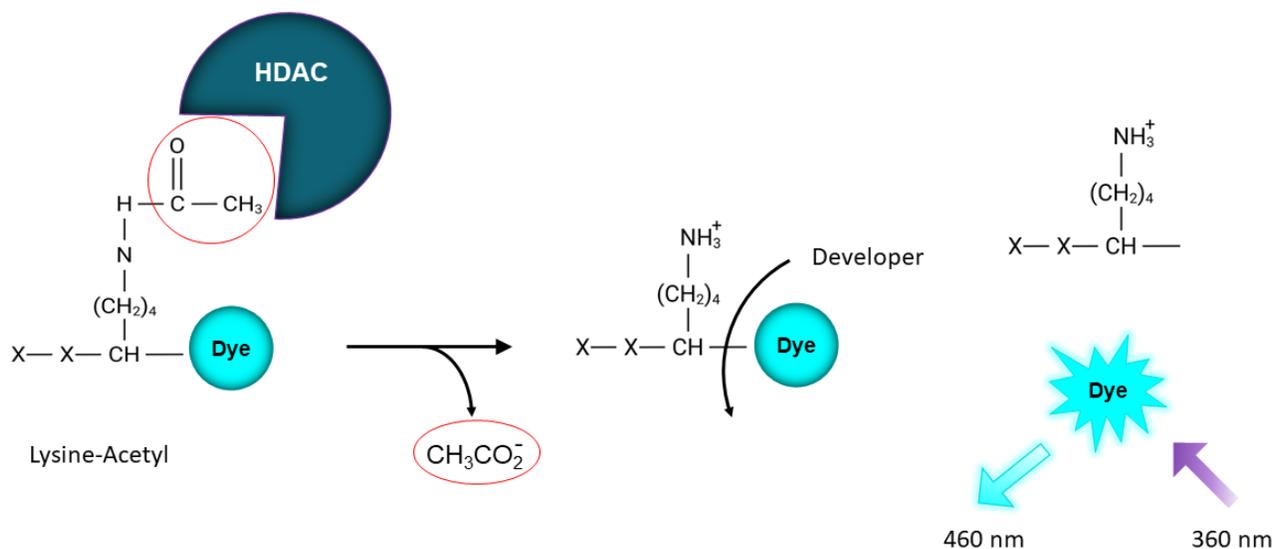


Figure 1: Illustration of the mechanism behind the HDAC2 Fluorogenic Assay Kit.

The fluorescence from dye molecules is quenched when bound to the peptide substrate. HDAC catalyzes the hydrolysis of the acetyl group from the lysine. Upon incubation with a developer solution specific for non-acetylated lysines, the dye is released and able to fluoresce ( $\lambda_{ex}$ =350-380 nm;  $\lambda_{em}$ =440-460 nm). Fluorescence is thus proportional to HDAC activity.

## Background

HDAC2, or histone deacetylase 2, is a Class I member of the histone deacetylase family which is involved in lysine deacetylation. Lysine acetylation/deacetylation is a dynamic process involved in the regulation of a variety of cellular functions, similarly to phosphorylation/dephosphorylation. Abnormal levels of HDAC2 contribute to cancer progression via multiple mechanisms and can be used as a biomarker, for instance in colorectal cancer (CRC). HDAC2 overexpression leads to AKT phosphorylation and hepatocarcinogenesis or regulates expression of K<sup>+</sup>-Cl<sup>-</sup> cotransporter-2 and pain in bone cancer. HDAC2 dysfunction has been linked to CRC, prostate cancer, bone, pancreatic and oral cancer, amongst others. Overexpression of HDAC2 contributes to chemoresistance to agents such as doxorubicin in CRC, and it has been shown that silencing HDAC2 can result in decreased activity of proteins such as AP-1 (activator protein 1), c-jun and c-fos. The use of combination therapy may be a potent therapeutical approach. HDAC-2 has also been linked to Parkinson's and Alzheimer's disease (AD). The development of new inhibitors specifically targeting HDAC2, in a disease-specific context, may open newer avenues for cancer and HDAC2-linked diseases.

## Applications

Study enzyme kinetics and screen small molecule inhibitors for drug discovery and high throughput screening (HTS) applications.

**Supplied Materials**

Catalog #	Name	Amount	Storage
50002-KC2	HDAC2, His-tag*	2 µg	-80°C
50037-KC25	5 mM Fluorogenic HDAC Substrate 3	25 µl	-80°C
50030	2x HDAC Developer (contains 2 µM Trichostatin A)	6 ml	-80°C
82748-KC100	200 µM Trichostatin A	100 µl	-20°C
50031-KC10	HDAC Assay Buffer	10 ml	-20°C
79685	Black, low binding microtiter plate	1	Room Temperature

\*The concentration of the protein is lot-specific and will be indicated on the tube.

**Materials Required but Not Supplied**

- 1 mg/ml BSA (bovine serum albumin) solution in distilled water
- Fluorimeter capable of excitation at  $\lambda=350-380$  nm and detection at  $\lambda=440-460$  nm
- Adjustable micropipettor and sterile tips
- Orbital shaker

**Storage Conditions**

This assay kit will perform optimally for up to **6 months** from date of receipt when the materials are stored as directed.

**Safety**

This product is for research purposes only and not for human or therapeutic use. This product should be considered hazardous and is harmful by inhalation, in contact with skin, eyes, clothing, and if swallowed. If contact occurs, wash thoroughly.

**Contraindications**

- The final concentration of DMSO in the assay should not exceed 1%.
- Compounds that are fluorescent may interfere with the results, depending on their spectral excitation and emission properties.
- It is recommended that the compound alone is tested to determine any potential interference of the compound with the assay results.

**Assay Protocol**

- All samples and controls should be performed in duplicate.
- The assay should include “Blank”, “Positive Control”, “Control Inhibitor” and “Test Inhibitor” conditions.
- We recommend maintaining the diluted protein on ice during use.
- For detailed information on protein handling please refer to Protein FAQs (bpsbioscience.com).

- We recommend using Trichostatin A as internal control. If not running a dose response curve for the control inhibitor, we recommend running the control inhibitor at 0.1X, 1X and 10X the IC<sub>50</sub> value shown in the validation data below.

1. Thaw **5 mM Fluorogenic HDAC Substrate 3**, **200 μM Trichostatin A** and **HDAC Assay Buffer**.
2. Prepare the **Inhibitor Control** by diluting 200 μM Trichostatin A to 1000X the IC<sub>50</sub> in 100% DMSO. Then dilute 10-fold in HDAC Assay Buffer (the DMSO amount is now 10%) and corresponds to 100X the IC<sub>50</sub> value (5 μl/ well). Using Diluent Solution prepare solutions at 1X and 10X the IC<sub>50</sub> value (5 μl/ well).
3. Dilute 25-fold the **5 mM Fluorogenic HDAC Substrate 3** with HDAC Assay Buffer (5 μl/well will be needed). This makes 200 μM HDAC Substrate 3.
4. Thaw **HDAC2** on ice. Briefly spin the tube to recover the full content.
5. Dilute HDAC2 to 1 ng/μl (5 μl/well) with HDAC Assay Buffer.
6. Prepare a **Master Mix** (35 μl/well): N wells x (30 μl of HDAC Assay Buffer + 5 μl of 1 mg/ml BSA).
7. Add 35 μl of Master Mix to every well.
8. Prepare the **Test Inhibitor** (5 μl/well): for a titration, prepare serial dilutions at concentrations 10-fold higher than the desired final concentrations. The final volume of the reaction is 50 μl.

8.1 If the Test Inhibitor is water-soluble, prepare serial dilutions of the inhibitor 10-fold more concentrated than the desired final concentrations in HDAC Assay Buffer.

For the positive and negative controls, use HDAC Assay Buffer (Diluent Solution).

**OR**

8.2 If the Test inhibitor is soluble in DMSO, prepare the test inhibitor at a concentration 100-fold higher than the highest desired concentration in 100% DMSO, then dilute the inhibitor 10-fold in HDAC Assay Buffer-to prepare the highest concentration of the 10-fold intermediate dilutions. The concentration of DMSO is now 10%.

Using HDAC Assay Buffer containing 10% DMSO to keep the concentration of DMSO constant, prepare serial dilutions of the Test Inhibitor at 10-fold the desired final concentrations.

For positive and negative controls, prepare 10% DMSO in HDAC Assay Buffer (vol/vol) so that all wells contain the same amount of DMSO (Diluent Solution).

*Note: The final concentration of DMSO should not exceed 1%.*

9. Add 5 μl of Test Inhibitor to each well labeled "Test Inhibitor".

10. Add 5  $\mu$ l of Diluent Solution to the "Positive Control" and "Blank" wells.
11. Add 5  $\mu$ l of 20  $\mu$ M Trichostatin A to the "Control Inhibitor" wells.
12. Add 5  $\mu$ l of HDAC Assay Buffer to the wells designated as "Blank".
13. Add 5  $\mu$ l of diluted HDAC to the wells designated "Positive Control", "Control Inhibitor" and "Test Inhibitor".
14. Preincubate the plate for 30 minutes at Room Temperature (RT) with gentle agitation.
15. Initiate the reaction by adding diluted Fluorogenic HDAC Substrate 3 (200  $\mu$ M).
16. Incubate at 37°C for 30 minutes.

Component	Blank	Positive Control	Control Inhibitor	Test Inhibitor
Master Mix	35 $\mu$ l	35 $\mu$ l	35 $\mu$ l	35 $\mu$ l
Test Inhibitor	-	-	-	5 $\mu$ l
Diluted Trichostatin (20 $\mu$ M)	-	-	5 $\mu$ l	-
Diluent Solution	5 $\mu$ l	5 $\mu$ l	-	-
HDAC Assay Buffer	5 $\mu$ l	-	-	-
Diluted HDAC2 (1 ng/ $\mu$ l)	-	5 $\mu$ l	5 $\mu$ l	5 $\mu$ l
Diluted Fluorogenic HDAC Substrate 3 (200 $\mu$ M)	5 $\mu$ l	5 $\mu$ l	5 $\mu$ l	5 $\mu$ l
<b>Total</b>	<b>50 <math>\mu</math>l</b>	<b>50 <math>\mu</math>l</b>	<b>50 <math>\mu</math>l</b>	<b>50 <math>\mu</math>l</b>

17. Add 50  $\mu$ l of **2x HDAC Developer** to each well.
18. Incubate at RT for 15 minutes.
19. Immediately read in a fluorimeter or a microplate reader capable of excitation at  $\lambda$ =350-380 nm and detection at  $\lambda$ =440-460 nm.
20. The "Blank" value should be subtracted from all other readings.

## Example Results

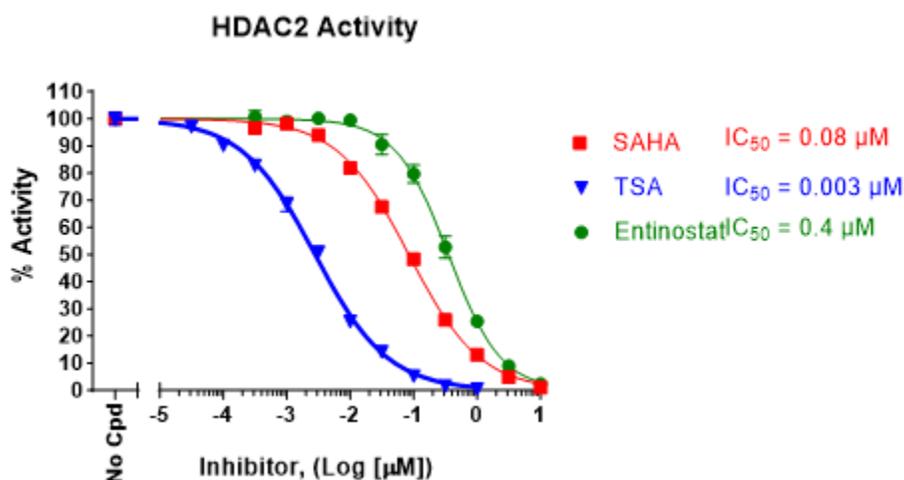


Figure 2: Inhibition of HDAC2 activity by the inhibitors SAHA, TSA (Trichostatin A) and Entinostat. HDAC2 activity was measured in the presence of increasing concentrations of SAHA (Cayman Chemicals #10009929), TSA and Entinostat (EMD Millipore #57625). The “Blank” value was subtracted from all other values. Results are expressed as the percent of control (activity in the absence of inhibitor, set at 100%).

Data shown is representative.

## Troubleshooting Guide

Visit [bpsbioscience.com/assay-kits-faq](https://bpsbioscience.com/assay-kits-faq) for detailed troubleshooting instructions. For lot-specific information and all other questions, please visit <https://bpsbioscience.com/contact>.

## References

- Santo L., *et al.*, 2012 *Blood*. 119(11):2579-89.  
 Bradner J.E., *et al.*, 2010 *Nat Chem Biol*. 6(3): 238-243.  
 Shetty M., *et al.*, 2021 *Pharmacological Research* 170: 105695.

## Related Products

Products	Catalog #	Size
HDAC2 Kinetic Assay Kit	53002	96 reactions
HDAC2, FLAG-Tag Recombinant	50052	50 µg
Anti-HDAC2 Monoclonal Antibody	25288	50 µg
Anti-HDAC1 Polyclonal Antibody	25287	50 µg
HDAC1 Fluorogenic Assay Kit	50061	96 reactions
HDAC3 Fluorogenic Assay Kit	50073	96 reactions

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